

NIST Special Publication 260 NIST SP 260-233r1

Value Assignment of Standard Reference Material® 1849b Infant/Adult Nutritional Formula I

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This publication is available free of charge from: <https://doi.org/10.6028/NIST.SP.260-233r1>

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August 2024

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Publication History

Approved by the NIST Editorial Review Board on 2024-08-23 Supersedes NIST Special Publication 260-233 (January 2023) https://doi.org/10.6028/NIST.SP.260-233

How to Cite this NIST Technical Series Publication

Phillips MM, Wood LJ, Barber CA, Scruggs BE, Sieber JR, Wood ESC, Yen JH, Yu LL (2024) Value Assignment of Standard Reference Material® 1849b Infant/Adult Nutritional Formula I. (National Institute of Standards and Technology, Gaithersburg, MD), NIST Special Publication 260-233r1. https://doi.org/10.6028/NIST.SP.260-233r1

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Abstract

The National Institute of Standards and Technology (NIST) Standard Reference Material® (SRM®) 1849b Infant/Adult Nutritional Formula I delivers certified or non-certified values for over 100 analytes. This material is intended to be used for the evaluation of methods for the determination of elements, amino acids, fatty acids, nucleotides, proximates, sugars, and vitamins and in this and similar matrices. The material was purchased pre-packaged from a commercial vendor. The measurement results used to assign values to the measurands were provided by NIST staff, the manufacturer, and/or interlaboratory comparison exercises. This document describes the material, measurement processes and results, and data analysis used to produce SRM 1849b.

Keywords

Amino Acids; Elements; Fatty Acids; Nucleotides; Proximates; Reference Material; Sugars; Vitamins.

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Acknowledgments

The authors thank Carolyn Vallone of the NIST Office of Reference Materials for providing the 1990 through 2002 sales history of SRM 1846 and Dave Duewer of the NIST Chemical Sciences Division for assistance in preparation of this report.

1. Introduction

Infant formula is one of the most highly regulated foods in the United States and around the world. The Infant Formula Act of 1980 (Public Law 96-359) requires that manufacturers test their products to make sure that nutrients fall within specified limits [1]. Beginning in 1996 with Standard Reference Material® (SRM®) 1846 Infant Formula, the National Institute of Standards and Technology (NIST) has provided reference materials for use in assuring the quality of infant formula-related nutrient measurements [2,3]. SRM 1846 was replaced with SRM 1849 Infant/Adult Nutritional Formula in July 2009 [4], followed by SRM 1849a Infant/Adult Nutritional Formula I (milk-based) in Dec 2011 [5]. A soy-containing material, SRM 1869 Infant/Adult Nutritional Formula II (milk/whey/soy-based), was introduced in 2018 [6]. In 2021, two materials were released to represent additional matrix types, including Reference Material (RM) 8260 Infant Nutritional Formula (hydrolyzed milk-based) [7] and RM 8261 Adult Nutritional Formula (high-protein) [8].

The SRM 1849 series of certified reference materials is located in sector 6 of the AOAC INTERNATIONAL food composition triangle [9,10] [\(Fig.](#page-17-1) 1). The information provided in this diagram is used by measurement laboratories in support of nutrition labeling. Laboratories needing an infant formula or similar material for the demonstration of method validity and accuracy when analyzing food products to generate data for nutrition labels can use the currently available edition of SRM 1849b. The SRM can also be used to test methodologies for food safety.

Fig. 1. NIST adaptation of the AOAC INTERNATIONAL food composition triangle. The white "+" depict the location of available food-matrix reference materials.

[Fig.](#page-18-0) 2 displays the sales history of the milk-based infant formula-related materials from the first sale of SRM 1846 in May 1996 to the sale of the last SRM 1849a unit in January 2020.

Fig. 2. Sales history of infant formula-related SRMs. The thick black line depicts the cumulative distribution of sales as a function of the order date, plotted using the "Units Sold" axis at the left of the plot. The thin blue line depicts the sales rate (the first derivative of the cumulative distribution), plotted using the "Sales Rate, Units per Year" axis to the right of the graph.

[Fig.](#page-18-1) 3 displays the proportion of sales to various countries or geographical regions over the past 24 years. Over the entire period, nearly 45 % of all sales have been within the USA and Canada, 24% to Europe, 23 % to Asia, 3 % to New Zealand, and the rest scattered elsewhere around the globe. The large proportion of sales to Germany since 2013 probably reflects European sales handled by a German-based NIST-licensed distributor.

Fig. 3. Location of customers for the infant formula-related materials. From left to right, the three charts display the proportion of sales to various countries or geographic regions from the first sale in 1996 through 12/31/2004, 1/1/2005 through 12/31/2012, and 1/1/2013 through the date of the last unit sold in 2020. Slices are shown for individual countries only when they purchased at least 2.5 % of the units sold during that interval. The area of the circle is proportional to the number of units sold during the interval.

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SRM 1849b Infant/Adult Nutritional Formula I (milk-based) is NIST's latest addition to the infant formula-related family of materials. As is true for all NIST food-matrix SRMs, SRM 1849b is intended for use as a primary control material when assigning values to in-house control materials and for validation of analytical methods for the measurement of nutrients in similar matrixes. SRM 1849b contains some nutrients at levels higher than compliant with Public Law 96-359 to provide broader material applicability while retaining its utility for use in measurement assurance within the infant formula industry.

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2. Material

Acquisition & Packaging

SRM 1849b is a milk-based hybrid infant/adult nutritional powder, prepared by a manufacturer of infant formula and adult nutritional products. A base liquid containing all constituents was conventionally heat processed, homogenized, and then spray-dried. The product was packaged into single-use nitrogen-flushed pouches, each containing 10 g of powder. Each unit of SRM 1849b contains 10 pouches. The inventory of the SRM is stored at NIST at –30 °C.

A total of 290 boxes (numbered 1 to 290) were produced and were assembled onto eight pallets (numbered 1 to 8) in approximate packaging order. Samples used to characterize the SRM were identified by pallet and box number.

2.2. Ingredients

[Table 1](#page-21-0) lists the ingredients used to prepare SRM 1849b and the approximate amounts of each per 1000 kg dry mass. All dairy and dairy-derived ingredients were certified by their providers as suitable for food grade use.

Table 1. Ingredients used to prepare SRM 1849b

2.3. Homogeneity

Homogeneity of the material was determined based on measurement data provided by the manufacturer. Eleven pouches were selected for analysis, two each from the boxes on pallets 1, 5, and 8 and one each from pallets 2, 3, 4, 6, and 7. The pouches from pallet 1, samples 1:1 and 1:2, represent the same approximate time point in the filling process as do those from pallet 8, samples 8:1 and 8:2. The pouches from pallet 5, samples 5:1 and 5:2, represent time points separated by about 20 min. Sample 5:2 represents the middle time point in the overall filling process.

The material manufacturer provided results for 129 measurands for all eleven pouches. These results confirmed that SRM 1849b is adequately homogenous across the production run and have been used to help assign quantitative values to many SRM 1849b components.

3. NIST Analyses

NIST WDXRF Analysis of Al, Ca, Cl, Cu, Fe, K, Mg, Mn, Na, P, Rb, S, and Zn

Mass fractions of aluminum (Al), calcium (Ca), chlorine (Cl), copper (Cu), iron (Fe), potassium (K), magnesium (Mg), manganese (Mn), sodium (Na), phosphorus (P), sulfur (S), rubidium (Rb), and zinc (Zn) were determined in SRM 1849b using WDXRF.

3.1.1. Materials

Six pouches of SRM 1849b, one each from boxes 1, 7, 109, 146, 182, and 290, were analyzed in duplicate. The pouches of SRM 1849b were selected using a stratified random sampling scheme based on packaging order. A single pouch of SRM 1849a was used as the quality assurance material, and four samples were prepared using the same procedure as for SRM 1849b. [Table 2](#page-23-3) lists the materials chosen from among the available powdered food and agricultural material SRMs for use as calibrants. These materials were prepared in the same way as the samples of SRM 1849a and SRM 1849b.

Table 2. SRMs used as WDXRF calibrants^a

^a COAs for these SRMs can be accessed at [https://shop.nist.gov.](https://shop.nist.gov/)

^b Expired and/or unavailable SRMs were internally verified and found to be fit-for-purpose for this analysis.

Blanks were prepared from microcrystalline cellulose from Alfa Aesar (Cat. No. A17730, lot 10200883; Ward Hill, MA, USA) and Whatman (Cat. No. CF11; Maidstone, UK). The cellulose powders were pressed into briquettes after drying with the food powders.

No chemical reagents are necessary for this analytical method.

3.1.2. Equipment

A Sartorius model LP1200S balance (Göttingen, DE) was used to weigh each specimen of powder prior to pressing to ensure approximately equivalent sample quantities among the various NIST SP 260-233r1 August 2024

materials. Exact weighing was not required since sample mass is not used in calculations of results. Samples are not diluted and minor variations in briquette thickness can be moderated by use of ratioing analyte signal to background intensity or Compton scatter.

Samples were pressed using a SPEX X-PRESS hydraulic press (model 3630; Metuchen, NJ, USA), operated in manual mode, with painted aluminum pressing caps (Somar, Cat. No. 3130; Reno, NV, USA). Briquettes were placed in double open-ended polyethylene sample cups (Cat. No. SC-4240, 40 mm; Premier Lab Supply, Port St. Lucie, FL, USA) fitted with Spectrolene polypropylene plastic film (Somar, 6 um thickness, Part No. 3506-33).

An Ultimate wavelength dispersive X-ray fluorescence spectrometer using the SuperQ 6.2b operating system (Malvern Panalytical, Model ZETIUM; Malvern, UK) was used to evaluate the samples.

3.1.3. Sample Preparation

Portions (4.5 g) of the sample, calibrant, and blank materials were weighed into glass containers (e.g., weighing bottles, Petri dishes and beakers) and stored in a desiccator over freshly regenerated silica gel for at least 21 d. To press briquettes, approximately 4.0 g of each dried powder was weighed onto weighing paper and poured into a 31 mm diameter steel pressing die containing an aluminum cap. A polished steel die pellet was inserted, along with the ram cylinder, and the die was pressed at 2 tons (2 000 kg) for 20 s under house vacuum using manual control of the press.

Briquettes were immediately placed into liquid sample cells fitted with 6 μ m polypropylene film, and the sample cells were placed into a second desiccator. The plastic film prevents loose powder from dropping into the spectrometer. When all briquettes had been prepared, the samples were loaded into the X-ray fluorescence spectrometer and measured.

3.1.4. Measurement Process

The X-ray spectrometer was used to measure the K-L2,3 characteristic X-ray lines of all elements in a helium environment with the generator operated at 4.0 kW of X-ray power. Background measurement and subtraction were made for Al, Ca, Cl, Cu, Fe, Mg, Mn, Na, Rb, and Zn. Liquid cell sample holders with a 31 mm inner diameter aperture were used to carry all briquettes in liquid cells into the spectrometer and to hold them in the measurement position. The mask between the sample and the collimator was set to view a 29 mm circular area of the sample, and the spinner was used. Counting times were chosen to obtain relative counting statistical errors ranging from 0.1 % to 1.0 % depending on the X ray count rate for each element and the number of measurements of background. The maximum measurement time per sample was approximately 14 min to avoid melting the polymer X-ray foil supporting the briquette and thermally induced swelling of the pressed briquettes. The WDXRF measurements were made in a pseudo-random order with unknown specimens interspersed with calibration standards.

Calibration curves were calculated in the SuperQ Analytical XRF software package (Malvern Panalytical). The generalized calibration algorithm is denoted in Equation 1

$$
C_i = D_i - L_{ik} \cdot C_k + E_i \frac{R_i}{R_{Rh}} \left(1 + \sum_j (\alpha_{ij} \cdot C_j) \right)
$$
 (1)

where *C* denotes mass fraction, *D* is the intercept, *L* is the line overlap factor, *E* is the inverse sensitivity, *R* is the gross or net WDXRF count rate, R_{Rh} is the gross count rate for Rh Compton scatter, and *α* is the absorption correction factor, with *i* indexing the analyte element, *k* the interfering element by line overlap, and *j* the interfering element by absorption. The Rh Compton scatter was used as an internal reference for Zn and Rb calibrations to compensate for effects of finite sample thickness on more energetic X-rays and some variations in matrix absorption effects. In a few cases (i.e., Ca, Cu, Fe, and Mn), the background intensity for Fe was used as an internal reference because the energy more closely matches that of these analyte lines and would be substituted for R_{Rh} in Equation [1.](#page-25-1)

The term containing the *α* factor for X-ray absorption was used for some elements to correct for variations in matrix X-ray absorption and enhancement effects [\(Table 3\)](#page-25-0). These *α* factors were calculated empirically based on the measured count rates of the X-ray lines from the interfering elements in cases where no mass fraction values reported for some analytes in the SRMs although some measurable peak was present. Selection of these absorption corrections factors was based on basic X-ray fluorescence theory of matrix effects and how well these factors improved the fit of the calibration points to the modeling equation.

Table 3. Absorption corrections for WDXRF measurements

A line overlap correction is used when the analyte line of interest significantly overlaps with some other first order fluoresced X-ray signal or higher order diffraction lines. In this analysis, an intensity-based line overlap correction was used to adjust the Na signal for the overlap of the Zn L-series lines.

For some analytes, selected calibrants were observed to be outliers and were excluded from the calibration [\(Table 4\)](#page-26-1). In some instances, exclusions were made automatically by the system software because the value of the analyte was provided at an informational level only. Other standards were excluded in instances where no value was provided on the COA. Calibrations for Cl, K, Mg, P and S were forced through the origin which effectively negates the influence of the blanks.

Table 4. SRMs excluded from use as WDXRF calibrants for selected analytes

3.1.5. Measurement Uncertainties

The standard uncertainty for each of the measurements, *uc*, combines three uncertainty components as described in Equation 2 and [Table 5.](#page-27-0)

$$
u_c = \sqrt{\frac{s^2}{n} + \frac{u_{\rm m}^2}{3} + (\bar{x}^2 \cdot \frac{u_{\rm m}^2}{3M^2})}.
$$
 (2)

Table 5. Uncertainty budget for WDXRF analysis

No error weighting: $u_m \approx |C_{\text{chem}} - C_{\text{calc}}| = RMS$

Linear error weighting:
$$
u_m \approx |C_{\text{chem}} - C_{\text{calc}}| = |\frac{RE \cdot W_0 + C_{\text{calc}}}{1 - RE} - C_{\text{calc}}|
$$
 (3)
Square root error weighting: $u_m \approx |C_{\text{chem}} - C_{\text{calc}}| \cong K \cdot \sqrt{C_{\text{calc}} + W_0}$

where *RMS*, *RE*, and *K* are the regression minimization factors for the respective weighting function and W₀ is a weighting constant. Weighting functions are employed to place higher significance within the curve fitting to different regions of the calibration curve (e.g., the low mass fraction extreme of the calibration versus the high mass fraction extreme). All calibrations used a square root error weighting except for S, K, and Rb which used no error weighting. The calibration process and determination of regression minimization factors are done in SuperQ software.

NIST ICP-MS Analysis of Cr, Mo, Se, and I

Mass fractions of chromium (Cr), molybdenum (Mo), selenium (Se), and iodine (I) were determined in SRM 1849b using ICP-MS.

3.2.1. Materials

Ten pouches of SRM 1849b, one each from boxes 1, 27, 58, 62, 104, 144, 166, 202, 260, and 290, were analyzed in duplicate. The pouches of SRM 1849b were selected using a stratified random sampling scheme based on packaging order. A single pouch of SRM 1849a was used as the quality assurance material, and four samples were prepared using the same procedure as for SRM 1849b.

Calibration solutions were prepared from SRM 3112a Chromium (Cr) standard Solution Lot No. 170630, SRM 3134 Molybdenum (Mo) Standard Solution Lot No. 130418, SRM 3140 Selenium (Se) Standard Solution Lot No. 100901, and SRM 3180 Iodide Anion (I⁻) Standard Solution Lot No. 110530. The internal standard was prepared from SRM 3102a Antimony (Sb) Standard Solution (Lot No. 149011). COAs for these SRMs are available at [https://shop.nist.gov.](https://shop.nist.gov/)

All samples were prepared using Optima grade acids from Fisher Scientific (Waltham, MA, USA). The iodine analyses also used scintillation grade Triton X-100 from Acros Organics (Waltham, MA, USA) and Optima grade NH4OH from Fisher Scientific. All dilute acid or base concentrations are expressed in volume fractions with respect to the concentrated acid or base.

3.2.2. Equipment

An Agilent Technologies 7500cs ICP-MS (Santa Clara, CA, USA), equipped with a Peltier-cooled, inert sample introduction system, was used to measure Cr, Mo, Se, I, and Sb in all solutions with read times per mass of 3 s and integration time of 0.1 s per point. Cr, Mo, and Se were measured in two runs; I was measured in three runs. For the Cr, Mo, and Se measurements H_2 was used as the collision gas to minimize polyatomic interferences. For the I measurements, the reaction cell mode was off.

A Mettler AT261 Delta Range analytical balance was used for weighing in the preparation of samples and standards. The balance is serviced and calibrated annually by Mettler. Prior to use, calibration is verified using standard masses ranging from 0.5 g to 20 g that are traceable to the SI.

3.2.3. Sample Preparation

Two 0.5 g aliquots were taken from each pouch of SRM 1849b and four 0.5 g aliquots were taken from one pouch of SRM 1849a and placed in Teflon microwave vessels. Ten procedural reagent blanks were also prepared along with the samples. To each vessel, 10 mL of concentrated $HNO₃$ and an aliquot of the Sb internal standard solution were added before microwave digestion. The

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Sb internal standard solution was prepared at 1000 ng/g Sb from SRM 3102a Antimony (Sb) Standard Solution with a final acid concentration of 1.5% $HNO₃$ (volume fraction).

3.2.3.1. Chromium, Molybdenum, and Selenium

To each vessel, 0.5 mL of the Sb internal standard solution was added prior to digestion. Samples were digested using MARSXpress vessels in a MARS5 Microwave Digestion (CEM Corporation, Matthews, NC, USA) system using 1600 W power at 85 %, a 20 min ramp time, temperature of 195 °C, and a 20 min hold time. After microwave digestion, solutions were transferred to PTFE beakers and heated on a hot plate with a surface temperature of approximately 180 °C until the volume was reduced to near dryness. Samples were transferred to polyethylene bottles and diluted to 30 g using 1.5 % $HNO₃$ (volume fraction).

The first aliquot of the pouch from box 290 provided no usable result due to a laboratory error. All results for Cr, Mo, and Se in SRM 1849b are based on the analysis of 19 validly prepared samples.

3.2.3.2. Iodine

To avoid loss of iodine in acidic solutions, a published digestion procedure was used [11]. To the SRM 1849a samples and each blank, 0.5 mL of the Sb solution was added; to the SRM 1849b samples, 0.8 mL was added. To all vessels, 0.4 mL of a 5 % volume fraction Triton X-100 solution was added as a surfactant. Samples were digested using CEM MARSXpress vessels in a CEM MARS5 Microwave Digestion system using the program described in [Table 6.](#page-29-3)

After microwave digestion, sample solutions were transferred to polyethylene bottles and diluted to 60 g by first adding 18 g of 18 MΩ·cm water followed by 30 g of a 7.5 % NH4OH solution (volume fraction). The diluted samples were tested to ensure that the pH was at or above pH 8. Ammonium hydroxide was used to immediately neutralize the acidic solutions.

3.2.4. Measurement Process

For determination of Cr, Mo, and Se, the method of standard additions was used for calibration. Iodine was determined using an external standard calibration approach.

3.2.4.1. Standard Additions: Cr, Mo, and Se

From each sample dilution, two aliquots were taken, with a spike added to one as described in [Table 7](#page-30-2) to the approximate analyte mass fractions.

		In	Added	Total in
Element	Standard	Solution	as Spike	Spiked Aliquot
Chromium (Cr)	SRM 3112a		b	11
Molybdenum (Mo)	SRM 3134	10	11	22
Selenium (Se)	SRM 3149		6.5	11.5

Table 7. Standards and approximate mass fractions for ICP-MS analysis, ng/g

Ten instrumental measurements were recorded and averaged for each sample aliquot and each spiked aliquot. Final mass fractions in SRM 1849b were calculated using the method of standard additions in which the analytical instrument is calibrated by measuring the increase in the analytical signal that occurs when a known amount of the analyte is added to the sample. This approach avoids multiplicative types of matrix interferences (enhancements or suppressions) since the calibrant is present within the same matrix as the sample. The method of standard additions can be used in any situation where an analyte and an internal standard can be quantitatively and homogeneously spiked into the sample.

The mass fraction of the analyte in the analyte in the sample, *F*_{sample}, is calculated using Equation 4:

$$
F_{\text{sample}} = R_{\text{u}} \left(\frac{\binom{m_{\text{sp}} F_{\text{sp}}}{m_{\text{sp}} - n_{\text{u}}} }{n_{\text{sp}} - n_{\text{u}}} \right) \left(\frac{m_{\text{solu}}}{m_{\text{sample}}} \right) \tag{4}
$$

where: F_{SD} mass fraction of the analyte in the spiking solution

*m*sample mass of sample that is present in the solution to be analyzed m_{solu} total mass of the sample solution after addition of the IS spike m_{sp} mass of the analyte spiking solution delivered to the solution *m*spsolu mass of the solution that will be spiked

*R*sp analyte/IS signal ratios for the spiked solution

R^u analyte/IS signal ratios for the unspiked solution.

3.2.4.2. Standard Additions with External Calibration: I

SRM 3180 *Iodide Anion (I-) Standard Solution* was used to prepare the iodine (I) spike and the standards for the external calibration curve for this analysis. From each sample dilution, two aliquots were taken, with a spike added to one as described i[n Table 8](#page-31-1) to the approximate analyte mass fraction.

The iodine mass fraction in SRM 1849b was calculated using external standard calibration since the Sb internal standard instrumental counts were often higher in the spiked sample than the unspiked sample. Limited laboratory access during the time of these experiments prevented prompt analysis immediately after sample preparation, which may have led to such an anomaly if Sb concentrations are more sensitive to the variations of pH over time than iodine concentrations. Ten instrumental measurements were recorded and averaged for each sample aliquot.

External calibration is accomplished by observing the instrument's responses, *y*, to different levels of the analyte, *x*. For iodine, the relationship was linear

$$
y = b_1 x + b_0, \tag{5}
$$

where *y* is the measured response, *x* is the mass of iodine (in ng/g) added to each point, b_1 is the sensitivity (slope), and b_0 is the value of y when $x = 0$ ng/g. This calibration curve is then used to obtain the mass, x_{pred} , of the analyte from a sample which produces an observed response y_{obs} from Equation 6:

$$
x_{\text{pred}} = (y_{\text{obs}} - b_0)/b_1 \tag{6}
$$

The constants b_1 and b_0 are determined by least squares regression on a set of n pairs of values $\{x_i, y_i\}$, and the result for the analysis is the value calculated for x_{pred} .

3.2.5. Measurement Uncertainties

The standard uncertainty associated with the mean value of each element was estimated as summarized in Equation 7 and [Table 9.](#page-32-1)

$$
u = \sqrt{s_{\text{sample}}^2 + s_{\text{blank}}^2 + u_{\text{S}}^2 + u_{\text{b1}}^2 + u_{\text{b2}}^2}
$$
 (7)

Table 9. Uncertainty budget for ICP-MS analysis

Approximate 95 % level of confidence expanded uncertainties for the mean values were estimated using Equation 8,

$$
U_{95\%} = k_{95\%}u\tag{8}
$$

where $k_{95\%}$ is the Student's *t* 95th percentile two-tailed expansion factor appropriate to the df associated with the standard uncertainty, estimated using the Welch-Satterthwaite formula [12, Section G.4.1].

NIST ICP-OES Analysis of Cu, Fe, Mg, and P

Mass fractions of magnesium (Mg), phosphorus (P), iron (Fe), and copper (Cu) were determined in SRM 1849b using ICP-OES.

3.3.1. Materials

The samples used for ICP-OES were those prepared and digested for ICP-MS measurements as described in Section 5.2.3.1.

Internal standard solutions were prepared from SRM 3148a Scandium (Sc) Standard Solution Lot No. 10070 and SRM 3124a Indium (In) Standard Solution Lot No. 110516. COAs for these SRMs are available a[t https://shop.nist.gov.](https://www.nist.gov/srm)

All samples were prepared using Optima grade acids from Fisher Scientific. Samples and acids were diluted using 1.5% HNO₃ (volume fraction). All dilute acid concentrations are expressed in volume fractions with respect to the concentrated acid.

3.3.2. Equipment

A Perkin-Elmer Optima 8300 Dual View ICP-OES (Waltham, MA, USA) was used to measure Cu, Fe, Mg, and P in all solutions as described in [Table 10.](#page-33-3)

A Mettler AT261 Delta Range analytical balance was used for weighing in the preparation of samples and standards. The balance is serviced and calibrated annually by Mettler. Prior to use, calibration is verified using standard masses ranging from 0.5 g to 20 g that are traceable to the SI.

3.3.3. Sample Preparation

The solutions used were those used for ICP-MS measurements described in Section 5.2.3.1. To each prepared solution, weighed aliquots containing 0.25 mL of a solution containing 100 µg/g In and 1 mL of a solution containing 100 μ g/g of Sc were added as internal standards to improve the precision of the instrumental measurements. The concentration of Sb in these solutions, approximately 16 ng/g as added for ICP-MS analysis, was below the estimated detection limit for ICP-OES and therefore Sb was not used as an internal standard.

3.3.4. Measurement Process

For determination of Cu, Fe, Mg, and P, the method of standard additions was used for calibration. Solutions were diluted and spiked to yield the approximate mass fractions listed in [Table 11.](#page-34-3) Analyte mass fractions were quantified by the method of standard additions (see Sectio[n 3.2.4.1\)](#page-30-0).

Table 11. Standards and approximate mass fractions for ICP-OES analysis, mg/kg

3.3.5. Measurement Uncertainties

See Section [3.2.5.](#page-31-0)

NIST LC-ICP-MS Analysis of Vitamin B¹² (Cyanocobalamin)

The mass fraction of CNCbl in SRM 1849b was determined using LC-ICP-MS [13] and single-point standard addition with use of an internal standard. Vitamin B_{12} exists in four forms: hydroxo-, methyl-, 5-deoxyadenosyl-, and cyano-cobalamin (CNCbl). In this work, all forms of vitamin B_{12} were converted to CNCbl before the measurement to increase the detection limit and permit SI traceability. By convention, all forms of the vitamin B_{12} cobalamins were reported as mass fraction of CNCbl.

3.4.1. Materials

Eight pouches of SRM 1849b, one each from boxes 1, 41, 157, and 290 and two each from boxes 76 and 182 were analyzed. The pouches of SRM 1849b were selected using a stratified random sampling scheme based on packaging order. Two pouches of SRM 1869 Infant/Adult Nutritional Formula II [\[6\]](#page-17-2) were used as the quality assurance material, and five samples were prepared using the same procedure as for SRM 1849b.

SRM 3113 Cobalt (Co) Standard Solution was used to prepare internal standards and to establish SI traceability of the USP Reference Standard Cyanocobalamin (Cat. 1152011, Lot F07440, Rockville, MD, USA).

HPLC grade methanol, HPLC grade acetonitrile, and ACS grade ethylenediamine tetraacetic acid (EDTA) diammonium salt were obtained from Fisher Scientific. Taka-diastase was obtained from Accurate Chemical (Carle Place, NY, USA). All other chemicals were of ACS grade, including sodium acetate trihydrate (NaOAc), glacial acetic acid (HOAc), and potassium cyanide (KCN ≥ 95 % purity). Locally prepared sub-boiling distilled water was used as a solvent.

3.4.2. Equipment

A Perkin-Elmer LC system equipped with a Peltier-cooled Series 200 autosampler and a Series 200 quaternary pump was coupled to a Perkin-Elmer Elan DRC II ICP-MS was used to measure CNCbl. Separation of CNCbl from cobalt (Co) and subsequent detection were accomplished using instrument parameters described in [Table 12.](#page-35-1)

System	Component	Description	
Perkin Elmer Series 200 LC	Column	Waters Atlantis T3 150 mm x 2.1 mm i.d.	
	Mobile phase	20 mmol/L EDTA in 25:75 methanol: water (volume fraction)	
	Flow rate	200 µL/min, isocratic	
	Injection volume	$25 \mu L$	
Perkin Elmer DRCII ICP-MS	RF power	1300 W	
	Nebulizer Gas Flow	1.04 mL/min	
	Sample introduction	Concentric nebulizer/cyclonic spray chamber	

Table 12. LC-ICP-MS parameters for the determination of cyanocobalamin

A Mettler model AT261 Delta Range analytical balance was used for weighing during the preparation of samples and standards. The balance is serviced and calibrated annually by Mettler. Prior to use, calibration of the balance was verified using standard masses ranging from 0.5 g to 50 g that are traceable to the SI. A Jouan model C312 centrifuge (Thermo Fisher Scientific, Waltham, MA, USA) was used to separate particulate matter from the aqueous phase of the samples. A Fisher Scientific Isotemp oven, model number 737F, was used for sample preparation. An Anton-Paar model DMA 35 density meter (Graz, AT) was used for density measurements.

3.4.3. Sample Preparation

Single 3 g aliquots from each pouch of SRM 1849b and five 3 g aliquots from the two pouches of SRM 1869 were accurately weighed into Kimax brand Class A 100 mL volumetric flasks. The samples from boxes 1 and 157 were prepared on day 1, boxes 41 and 290 on day 2, and boxes 76 and 182 on day 3. Three procedural blanks were processed similarly, with a procedural blank and at least one sample of SRM 1869 included on each day of SRM 1849b sample preparation. To each flask, an aliquot of 25 mL of de-ionized water was added and the contents were swirled until the sample powder was dissolved. A 1 mL aliquot of 6 % (mass fraction) taka-diastase was added to each flask, and the contents were mixed by vortexing. The flask was placed in a box devoid of light for 40 min to allow the reaction of the contents to reach completion [14].

At the completion of the reaction, a 30 mL aliquot of 0.25 mol/L NaOAc buffer at pH 4.5 was added to each flask, and the contents were mixed by swirling. A 1 mL aliquot of 1 % (mass fraction) KCN was added to each flask, and samples were heated at 105 °C in an oven for 70 min. After heating, the samples were immediately cooled in an ice bath to near room temperature. The contents of each flask were diluted to volume with water and transferred to two 50 mL polyethylene tubes. Tubes were centrifuged for 30 min at 367 rad/s (3500 rpm) in a Jouan centrifuge. After the centrifugation, the supernatants from each tube were filtered through Whatman 2V filter paper and combined into a 125 mL Erlenmeyer flask.

A Maxi-Clean C18 900 mg SPE cartridge from S*PURE (Part No. 20942/5122344; Singapore) was attached to a 20 mL syringe. Before use, the SPE cartridges were conditioned with 20 mL acetonitrile and rinsed with 10 mL water by gently pressing the piston of the syringe. The
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conditioned SPE cartridges with the 20 mL syringe barrels were inserted onto the stopcock of the vacuum manifold. An 80 mL aliquot of each sample filtrate was passed through the cartridge. The exact volume of each aliquot was determined by the mass of the aliquot and the density of the solution measured with a separate portion of the filtrate. The effluent was monitored to exit the SPE cartridges at approximately 40 drops/min to 100 drops/min. After all the sample filtrate passed through the cartridges, the cartridges were air-dried by pulling vacuum until no more effluent was observed, and the stopcock was closed. A pre-weighed 15 mL Falcon tube was placed under the cartridge, and a 5 mL aliquot of 30 % (volume fraction) acetonitrile in water was added to the syringe. Vitamin B_{12} (as CNCbI) in the SPE cartridges was eluted into the Falcon tubes, assisted by gently pressing the piston. An aliquot of 0.2 g solution containing 90 ng/g Co was added to each eluate as an internal standard, and the mass of each tube was weighed again to determine the mass of the contents by the difference. Each sample was filtered through a 0.45 µm Nylon filter. A 0.6 g aliquot of each subsample and a 0.1 g aliquot of a solution containing 15 µg/kg Co as CNCbl were transferred into a 0.75 mL polypropylene vial to constitute a spiked sample for the purpose of quantification by the method of standard addition. A 0.6 g aliquot of each subsample and 0.1 g water were transferred into a separate 0.75 mL polypropylene vial to constitute an unspiked sample for the measurement.

3.4.4. Measurement Process

The mass fractions of CNCbl in the spiked and the unspiked samples were measured with LC--ICP-MS using the separation and spectrometric parameters listed in [Table 12](#page-35-0) by the method of standard additions (see Section [3.2.4.1\)](#page-30-0). Free Co from the internal standard and Co from CNCbl were measured at 59 Da in the standard mode of the DRCII ICP-MS instrument. All samples were measured in a continuous run spanning two days. Two analytical runs were conducted on the samples with 10 µL injection volume per sample in the first run and 25 µL injection volume per sample in the second run. The results for 10 μ L and 25 μ L injection volumes per sample were found statistically equivalent; however, the results using the 25 µL injection volume per sample have a better signal-to-background ratio and were used for the quantitative analysis. [Fig.](#page-37-0) 4 displays a typical chromatogram of an SRM 1849b sample spiked with Co as the internal standard.

Fig. 4. Exemplar LC-ICP-MS chromatogram for cyanocobalamin in SRM 1849b

3.4.5. Measurement Uncertainty

The standard uncertainty of these LC-ICP-MS measurements is estimated using Equation 9, with the components and their estimated values as described in [Table 13.](#page-37-1)

$$
u = \sqrt{u_{\rm rep}^2 + B_1^2 + B_2^2 + B_3^2 + B_4^2 + B_5^2} \,. \tag{9}
$$

NIST ID LC-MS Analysis of Free Carnitine and Total Choline

The mass fractions of free carnitine and total choline in SRM 1849b were determined using microwave-assisted hydrolysis and ID LC-MS [15].

3.5.1. Materials

Six pouches of SRM 1849b, one each from boxes 1, 36, 61, 149, 282, and 290 were analyzed. The pouches of SRM 1849b were selected using a stratified random sampling scheme based on packaging order. Two pouches of SRM 1849a Infant/Adult Nutritional Formula I (milk-based) [\[5\]](#page-17-0) were used as the quality assurance material, and six samples were prepared using the same procedure as for SRM 1849b.

Carnitine hydrochloride (Cat. C9500-5g, Lot 000133675) and choline bitartrate (Cat. C1629-100g, Lot 011M2016V) were obtained from Sigma-Aldrich (St. Louis, MO, USA) and were used in the preparation of calibration solutions. Purity of the calibrant materials was assessed using quantitative proton NMR (q^1 H-NMR) to provide metrological SI-traceability of the measured values. The internal standards, choline chloride trimethyl-*d⁹* (Cat. D-2142, Lot M280P1) and DLcarnitine HCl trimethyl-*d⁹* (Cat. D-5780, Lot #DE-245), were obtained from C/D/N Isotopes (Pointe-Claire, QC, Canada).

Hydrochloric acid, ACS grade was purchased from Taylor Scientific (Cat. H2620P-10, Lot 2016031714, St. Louis, MO, USA), ammonium formate (Cat. 70221, Lot BCCB6321) was purchased from Sigma-Aldrich, and sodium hydroxide pellets (Cat. S318-500, Lot 172340) were purchased from Fisher Chemical. HPLC grade water and acetonitrile were used for calibrant and sample preparation, as well as for preparation of LC-MS mobile phases.

3.5.2. Equipment

An Agilent 1200 Series LC system equipped with a binary pump, degasser, autosampler, and column compartment and an Agilent 6130 Quadrupole LC-MS were used to measure carnitine and choline. Separation of carnitine and choline from matrix components and subsequent detection were accomplished using instrument parameters described in [Table 14.](#page-39-0)

Table 14. LC-MS parameters for the determination of carnitine and choline

A Mettler model XPR205 analytical balance was used for weighing during the preparation of samples and standards. The balance is serviced and calibrated annually by Mettler. A Microwave Assisted Reaction System (MARS) from CEM Corporation (Matthews, NC, USA) was used for sample preparation. A Beckman Coulter Allegra model X-14R centrifuge (Brea, CA, USA) was used to separate particulate matter from the aqueous phase of the samples.

3.5.3. Sample Preparation

Triplicate 1 g aliquots from each pouch of SRM 1849b and SRM 1849a were accurately weighed into 50 mL polyethylene centrifuge tubes. One sample from each box was prepared on each of 3 days. To each tube, aliquots of 660 μL of carnitine-*d9* solution and 574 μL of choline-*d9* solution, as well as 30 mL of 1 mol/L hydrochloric acid solution. The contents of each tube were mixed by vortexing and transferred into a Teflon Xpress vessel (CEM Corporation, Matthews, NC, USA) designed for microwave extraction. Samples were heated to 110 °C over 15 min and held at 110 °C for 30 min. After cooling to room temperature, samples were individually transferred to clean 50 mL polyethylene tubes. The pH of each sample was adjusted to pH 3.8 to pH 4.0 using 50 % (w/v) sodium hydroxide and confirmed with litmus paper. Samples were centrifuged at 3000 rpm for 15 min and approximately 2 mL to 4 mL of the supernatant was filtered through a 0.45 μm regenerated cellulose syringe filter. Filtered samples were diluted 200-fold by combining 50 μL of supernatant with 10 mL water. Approximately 1 mL of the final diluted sample was transferred to an HLPC vial and stored under refrigeration (2 °C to 8 °C) until analysis by LC-MS.

3.5.4. Measurement Process

The mass fractions of carnitine and choline in the prepared samples were measured with LC-MS using the separation and spectrometric parameters listed in [Table 14](#page-39-0) through the method of isotope dilution. The MS peak area was manually integrated and recorded for each chromatographic peak, and a response factor for each the calibrants was calculated as described in equation 10.

$$
response\ factor = \frac{peak\ area\ (analyte)}{peak\ area\ (internal\ standard)} \times \frac{mass\ (internal\ standard)}{mass\ (analyte)}
$$
 (10)

The average response factors were calculated across independently prepared calibrants injected on the three different days. The average response factor was then used to calculate the mass fraction of carnitine and choline in each sample, as described in equation 11.

analyte mass fraction =
$$
\frac{peak\ area\ (analyte)}{peak\ area\ (internal\ standard)} \times \frac{mass\ (internal\ standard)}{response\ factor} \times \frac{1}{mass\ (sample)}
$$
 (11)

[Fig.](#page-40-0) 5 displays a typical chromatogram of an SRM 1849b sample spiked with isotopically labeled carnitine and choline as the internal standards.

Fig. 5. Exemplar ID LC-MS chromatogram for carnitine and choline in SRM 1849b

4. Interlaboratory Studies

Results from four NIST-coordinated ILS were used to help assign quantitative values to many SRM 1849b components: HAMQAP Exercises 5 and 6 and FNSQAP Exercises 1 and 2. HAMQAP was established in collaboration with the NIH ODS in 2017 to enable laboratories to improve the accuracy of measurements in samples that represent human intake (e.g., foods, dietary supplements, tobacco) and samples that represent human metabolism (e.g., blood, serum, plasma, urine) for demonstration of proficiency and/or compliance with various regulations. FNSQAP was launched in 2021 following reorganization of NIST QAPs to better serve various stakeholder groups, but with the same intention as HAMQAP, to help laboratories improve the accuracy of measurements in food samples. Participation in HAMQAP and FNSQAP is voluntary and anonymous.

Samples of SRM 1849b were tested for the following measurands via HAMQAP:

- Exercise 5, Spring 2020, prefix "E": thiamine (vitamin B_1), riboflavin (vitamin B_2), niacinamide (vitamin B₃), pantothenic acid (vitamin B₅), pyridoxine (vitamin B₆), ergocalciferol (vitamin D_2), and cholecalciferol (vitamin D_3) [16].
- • Exercise 6, Spring 2021, prefix "F": chromium, chlorine, iodine, molybdenum, selenium, retinyl acetate (vitamin A), retinyl palmitate (vitamin A), total retinol (vitamin A), biotin (vitamin B7), ascorbic acid (vitamin C), α-tocopherol (vitamin E), total α-tocopherol (vitamin E), α-tocopherol acetate (vitamin E), δ-tocopherol (vitamin E), γ-tocopherol (vitamin E), ash, calories, carbohydrates, fat, protein, and solids [17].

Samples of SRM 1849b were tested for the following measurands via FNSQAP:

- • Exercise 1, Summer 2021, prefix "A": calcium, sodium, iron, potassium, folic acid (vitamin B9), and vitamin K (phylloquinone) [18].
- Exercise 2, Winter 2022, prefix "B": choline, carnitine, β -carotene, lutein, lycopene, arachidonic acid (ARA), docosahexaenoic acid (DHA) [19].

Results from laboratory participants in each ILS are anonymized and are identified with a unique code consisting of an alphabetic study-specific prefix and an arbitrary numeric index ranging from one to a maximum equal to the number of participating laboratories in the exercise.

While all the results reported to the HAMQAP and FNSQAP ILS are presented in the following sections, results identified as non-representative using standard outlier detection methods are not used in value assignment. Such values are identified with a corresponding footnote in each table.

5. Results and Discussion

Results provided by the manufacturer, NIST, and/or by collaborating laboratories participating in one of the HAMQAP or FNSQAP ILS will be presented in a single table that displays all relevant results using the format outlined in [Table 15.](#page-42-0)

Table 15. Exemplar mass fraction results, mg/kg

- ^a The sample identifier for the manufacturer-supplied results concatenates the pallet and sample indices.
- b The arithmetic average of the sample replicates.</sup>
- c The standard deviation of the sample replicates.
- d The number of quantitative results available, either single values (for the manufacturer) or sample means (of two replicates for NIST results or three replicates for ILS results).
- ^e The arithmetic average of the (single or sample mean) quantitative results.
 f The standard deviation of the (single or sample mean) quantitative results
- The standard deviation of the (single or sample mean) quantitative results.
- g The pooled sample replicate standard deviation (effectively the average sample replicate SD).</sup>
- h The measurement uncertainty reported by the NIST analyst, where available.</sup>

When sufficient quantitative results are available, the data are also presented graphically as exemplified in [Fig.](#page-43-0) 6. The manufacturer's results are solid black circles, NIST results are various symbols with error bars representing one standard deviation above and below the mean, and ILS results are presented in boxplot format. The width of the box is proportional to the square root of the number of quantitative values; top, middle, and bottom lines of the box represent the 75th, $50th$, and $25th$ percentiles; and the whiskers span from the 2.5th to the 97.5th percentile. Results are ordered by the production sequence determined by the pallet and box index of the sampled pouch.

Fig. 6. Exemplar multi-source mass fraction as a function of production sequence

When only the manufacturer-provided results are available for a family of related measurands (e.g., amino acids), results are presented in tabular form using the format in [Table 16.](#page-43-1)

Table 16. Exemplar manufacturer-only mass fraction results, mg/kg

- ^a The sample identifier for the manufacturer-supplied results concatenates the pallet and sample indices. The sample results are presented in the order of analysis.
- b The arithmetic average of the eleven results, one per pouch analyzed.
- c The standard deviation of the eleven results.

Elements

5.1.1. Aluminum (Al)

The manufacturer electrothermal AAS [20] and NIST WDXRF measurement results for aluminum (Al) are summarized in [Table 17.](#page-44-0) The Al mass fraction is close to or below the limit of quantification for both methods.

Table 17. Summary of results for aluminum (Al), mg/kg

^a Identified as a non-representative value; excluded from statistical analysis.

5.1.2. Calcium (Ca)

The manufacturer ICP-OES [\[26\]](#page-62-0), NIST WDXRF, and FNSQAP [\[18\]](#page-41-0) measurement results for calcium (Ca) are summarized in [Table 18.](#page-45-0) [Fig.](#page-46-0) 7 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 18. Summary of results for calcium (Ca), mg/kg

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 7. Calcium (Ca) mass fraction as a function of production sequence

5.1.3. Chlorine (Cl)

The manufacturer potentiometric [21-23], NIST WDXRF, and HAMQAP [\[17\]](#page-41-1) measurement results for chlorine (Cl) or, equivalently, chloride (Cl⁻), are summarized in [Table 19.](#page-47-0) [Fig.](#page-47-1) 8 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 8. Chlorine (Cl) mass fraction as a function of production sequence

5.1.4. Chromium (Cr)

The manufacturer ICP-MS [24], NIST ICP-MS, and HAMQAP [\[17\]](#page-41-1) measurement results for chromium (Cr) are summarized in [Table 20.](#page-48-0) [Fig.](#page-48-1) 9 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 20. Summary of results for chromium (Cr), mg/kg

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 9. Chromium (Cr) mass fraction as a function of production sequence

5.1.5. Copper (Cu)

The manufacturer ICP-OES [\[26\]](#page-62-0), NIST WDXRF, and NIST ICP-OES measurement results for copper (Cu) are summarized in [Table 21.](#page-49-0) [Fig.](#page-49-1) 10 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 21. Summary of results for copper (Cu), mg/kg

Fig. 10. Copper (Cu) mass fraction as a function of production sequence

5.1.6. Fluorine (F)

The manufacturer ion-selective electrode measurement results for fluoride (F-), reported as fluorine (F), are summarized i[n Table 22.](#page-50-0) [Fig.](#page-50-1) 11 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 22. Summary of Results for Fluorine (F), mg/kg

Fig. 11. Fluorine (F) mass fraction as a function of production sequence

5.1.7. Iodine (I)

The manufacturer ICP-MS [25], NIST ICP-MS, and HAMQAP [\[17\]](#page-41-1) measurement results for iodine (I) are summarized in [Table 23.](#page-51-0) [Fig.](#page-51-1) 12 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 23. Summary of results for iodine (I), mg/kg

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 12. Iodine (I) mass fraction as a function of production sequence

5.1.8. Iron (Fe)

The manufacturer ICP-OES [\[26\]](#page-62-0), NIST WDXRF, NIST ICP-OES, and FNSQAP [\[18\]](#page-41-0) measurement results for iron (Fe) are summarized in [Table 24.](#page-52-0) [Fig.](#page-53-0) 13 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

^a The suspected reason for this higher data point is failure of proper washout as this sample was run directly after the sample calibration curve. Limited laboratory access prevented rerunning of particular sample to verify that this was the cause of the higher value.

Fig. 13. Iron (Fe) mass fraction as a function of production sequence

5.1.9. Magnesium (Mg)

The manufacturer ICP-OES [\[26\]](#page-62-0), NIST WDXRF, and NIST ICP-OES measurement results for magnesium (Mg) are summarized in [Table 25.](#page-54-0) [Fig.](#page-54-1) 14 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 25. Summary of results for magnesium (Mg), mg/kg

Fig. 14. Magnesium (Mg) mass fraction as a function of production sequence

5.1.10. Manganese (Mn)

The manufacturer ICP-OES [\[26\]](#page-62-0) and NIST WDXRF measurement results for manganese (Mn) are summarized in [Table 26.](#page-55-0) [Fig.](#page-55-1) 15 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 26. Summary of results for manganese (Mn), mg/kg

Fig. 15. Manganese (Mn) mass fraction as a function of production sequence

5.1.11. Molybdenum (Mo)

The manufacturer ICP-MS [\[24\]](#page-48-2), NIST ICP-MS, and HAMQAP [\[17\]](#page-41-1) measurement results for molybdenum (Mo) are summarized in [Table 27.](#page-56-0) [Fig.](#page-56-1) 16 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 27. Summary of results for molybdenum (Mo), mg/kg

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 16. Molybdenum (Mo) mass fraction as a function of production sequence

5.1.12. Phosphorus (P)

The manufacturer ICP-OES [\[26\]](#page-62-0) and NIST WDXRF and ICP-OES measurement results for phosphorus (P) are summarized in [Table 28.](#page-57-0) [Fig.](#page-57-1) 17 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer		NIST WDXRF						NIST ICP-OES					
ID	Value	Box	Rep ₁	Rep ₂	Mean	SD		Box	Rep ₁	Rep ₂	Mean	SD	
1:1	3620	1	3840	3745	3792	67		1	4002	3587	3795	293	
2:1	3500	7	3805	3752	3778	37		27	3931	3794	3863	97	
3:1	3610	109	3791	3864	3827	52		58	3897	3792	3845	74	
4:1	3650	146	3773	3808	3790	25		62	3970	3885	3928	60	
5:1	3760	182	3796	3833	3814	26		104	3773	3627	3700	103	
5:2	3560	290	3748	3855	3802	76		144	3733	3627	3680	75	
6:1	3660			N:	6			166	3896	3854	3875	30	
7:1	3630	3801 Mean, Pooled SD: 51						202	3868	3723	3796	103	
8:1	3650	SD: 18						260	3768	3977	3873	148	
1:2	3580			U:	171			290	3993		3993		
8:2	3670									N:	10		
N:	11									Mean, Pooled SD:	3821	130	
Mean:l	3626									SD:	135		
SD:l	67									U:	108		

Table 28. Summary of results for phosphorus (P), mg/kg

Fig. 17. Phosphorus (P) mass fraction as a function of production sequence

5.1.13. Potassium (K)

The manufacturer ICP-OES [\[26\]](#page-62-0), NIST WDXRF, and FNSQAP [\[18\]](#page-41-0) measurement results for potassium (K) are summarized in [Table 29.](#page-58-0) [Fig.](#page-59-0) 18 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 29. Summary of results for potassium (K), mg/kg

Fig. 18. Potassium (K) mass fraction as a function of production sequence

5.1.14. Rubidium (Rb)

The NIST WDXRF measurement results for rubidium (Rb) are summarized in [Table 30.](#page-60-0) [Fig.](#page-60-1) 19 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 30. Summary of results for rubidium (Rb), mg/kg

Fig. 19. Rubidium (Rb) mass fraction as a function of production sequence

5.1.15. Selenium (Se)

The manufacturer ICP-MS [\[24\]](#page-48-2), NIST ICP-MS, and HAMQAP [\[17\]](#page-41-1) measurement results for selenium (Se) are summarized in [Table 31.](#page-61-0) [Fig.](#page-61-1) 20 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 31. Summary of results for selenium (Se), mg/kg

Mean, Pooled SD: 0.795 0.063 SD: 0.096

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 20. Selenium (Se) mass fraction as a function of production sequence

5.1.16. Sodium (Na)

The manufacturer ICP-OES [26-28], NIST WDXRF, and FNSQAP [\[18\]](#page-41-0) measurement results for sodium (Na) are summarized i[n Table 32.](#page-62-1) [Fig.](#page-63-0) 21 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 32. Summary of results for sodium (Na), mg/kg

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 21. Sodium (Na) mass fraction as a function of production sequence

5.1.17. Sulfur (S)

The NIST WDXRF measurement results for sulfur (S) are summarized in [Table 33.](#page-64-0) [Fig.](#page-64-1) 22 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 33. Summary of results for sulfur (S), mg/kg

Fig. 22. Sulfur (S) mass fraction as a function of production sequence

5.1.18. Zinc (Zn)

The manufacturer ICP-OES [\[26\]](#page-62-0) and NIST WDXRF measurement results for zinc (Zn) are summarized in [Table 34.](#page-65-0) [Fig.](#page-65-1) 23 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 34. Summary of results for zinc (Zn), mg/kg

Fig. 23. Zinc (Zn) mass fraction as a function of production sequence

Vitamins and Related Measurands

5.2.1. Vitamin A Acetate (Retinyl Acetate)

The manufacturer LC-Abs [29] and HAMQAP [\[17\]](#page-41-1) measurement results for vitamin A acetate (retinyl acetate) are summarized in [Table 35.](#page-66-0) [Fig.](#page-66-1) 24 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 24. Vitamin A acetate (retinyl acetate) mass fraction as a function of production sequence

a

5.2.2. Vitamin A Palmitate (Retinyl Palmitate)

The manufacturer LC-Abs [\[29\]](#page-66-2) and HAMQAP [\[17\]](#page-41-1) measurement results for vitamin A palmitate (retinyl palmitate) are summarized in [Table 36.](#page-67-0) [Fig.](#page-67-1) 25 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Identified as a non-representative value; excluded from statistical analysis.

Fig. 25. Vitamin A palmitate (retinyl palmitate) mass fraction as a function of production sequence

5.2.3. Total Vitamin A (Retinol)

The manufacturer LC-Abs [\[29\]](#page-66-2) and HAMQAP [\[17\]](#page-41-1) measurement results for total vitamin A (retinol) are summarized in [Table 37.](#page-68-0) [Fig.](#page-68-1) 26 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 37. Summary of results for total vitamin A (retinol), mg/kg

a Identified as a non-representative or non-quantitative value; excluded from statistical analysis.

Fig. 26. Total vitamin A (retinol) mass fraction as a function of production sequence

5.2.4. Vitamin B¹ (Thiamine)

The manufacturer LC-FL and HAMQAP [\[16\]](#page-41-2) measurement results for vitamin B_1 (thiamine) are summarized in [Table 38.](#page-69-0) [Fig.](#page-69-1) 27 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer		HAMQAP Exercise 5								
Value ID	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Method			
1:1 14.48	E002	16.22	16.19	16.21	16.21	0.02	ILC-MS			
2:1 14.40	E005	14.00	14.00	14.00	14.00	0.00	LC-Abs or PDA			
3:1 14.48	E007	14.00	14.00	14.00	14.00	0.00	LC-Abs or PDA			
4:1 14.48	E010	18.66	18.24	18.44	18.45	0.21	LC-Abs or PDA			
5:1 14.48	E012	3.75 ^a	3.68 ^a	3.4 ^a			LC-Abs or PDA			
5:2 14.48	E023	13.30	13.60	13.90	13.60	0.30	LC-MS/MS			
6:1 14.48	E030	16.80	18.70	22.80	19.43	3.07	LC-Abs or PDA			
14.48 7:1	E040	9.90	10.40	7.70	9.33	1.44	ILC-MS			
8:1 14.56	E041	14.30	15.20	14.50	14.67	0.47	lLC-Abs or PDA			
1:2 14.48	E042	12.53	12.73	12.50	12.59	0.12	LC-MS			
8:2 14.56				N:	9					
N:111				Mean, Pooled SD:	14.70	1.14				
Mean: 14.48				SD:	3.05					
SD:l 0.04										

Table 38. Summary of results for vitamin B¹ (thiamine), mg/kg

a Identified as a non-representative value; excluded from statistical analysis.

Fig. 27. Vitamin B¹ (thiamine) mass fraction as a function of production sequence

5.2.5. Vitamin B² (Riboflavin)

The manufacturer LC-FL and HAMQAP [\[16](#page-41-2)] measurement results for vitamin B_2 (riboflavin) are summarized in [Table 39.](#page-70-0) [Fig.](#page-70-1) 28 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer		HAMOAP Exercise 5								
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods		
1:1	16.9	E002	16.37	16.96	16.39	16.57	0.33	LC-MS		
2:1	16.9	E005	20.00	20.00	20.00	20.00	0.00	LC-Abs or PDA		
3:1	17.0	E010	17.86	17.79	17.99	17.88	0.10	LC-Abs or PDA		
4:1	17.0	E012	3.66 ^a	3.45°	3.39 ^a			LC-Abs or PDA		
5:1	16.9	E013	$<$ 20 $^{\circ}$	$<$ 20 $^{\circ}$	$<$ 20 $^{\circ}$			LC-Abs or PDA		
5:2	16.7	E023	15.20	15.30	15.40	15.30	0.10	LC-MS/MS		
6:1	17.0	E030	16.30	18.70	16.90	17.30	1.25	LC-Abs or PDA		
7:1	16.9	E040	19.70	15.50	18.00	17.73	2.11	LC-MS		
8:1	16.9	E041	12.60	12.00	10.80	11.80	0.92	LC-Abs or PDA		
1:2	17.0	E042	5.345^a	5.117 ^a	5.245°			LC-MS		
8:2	17.0	E047	15.00	15.00	15.00	15.00	0.00	LC-Abs or PDA		
N:	11				N:	8				
Mean:	16.93				Mean, Pooled SD:	16.45	0.94			
SD:	0.09				SD:	2.45				

Table 39. Summary of results for vitamin B² (riboflavin), mg/kg

a Identified as a non-representative or non-quantitative value; excluded from statistical analysis.

Fig. 28. Vitamin B² (riboflavin) mass fraction as a function of production sequence

5.2.6. Vitamin B³ (Niacinamide)

The manufacturer LC-MS/MS and HAMQAP $[16]$ measurement results for vitamin B_3 (niacinamide) are summarized in [Table 40.](#page-71-0) [Fig.](#page-71-1) 29 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

	Manufacturer	HAMOAP Exercise 5								
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods		
1:1	112.0	E002	112.03	110.65	111.99	111.56	0.79	ILC-MS		
2:1	102.0	E005	100.00	101.00	101.00	100.67	0.58	lLC-Abs or PDA		
3:1	108.0	E007	99.95	97.79	97.89	98.54	1.22	lLC-Abs or PDA		
4:1	103.0	E010	111.32	111.09	110.88	111.10	0.22	ILC-Abs or PDA		
5:1	111.0	E013	$<$ 200 $^{\circ}$	$<$ 200 $^{\circ}$	$<$ 200 $^{\circ}$			LC-Abs or PDA		
5:2	112.0	E023	112.00	112.00	110.00	111.33	1.15	llC-MS/MS		
6:1	109.0	E030	126.00	139.00	127.00	130.67	7.23	ILC-Abs or PDA		
7:1	111.0	E040	165.00	140.40	168.30	157.90	15.24	ILC-MS		
8:1	105.0	E042	110.20	114.80	114.60	113.20	2.60	ILC-MS		
1:2	115.0	E047	340 ^a	340 ^a	350 ^a			LC-Abs or PDA		
8:2	99.1				N:	8				
N:	11				Mean, Pooled SD:	116.87	6.08			
Mean:	107.9				SD:	19.19				
SD:	5.0									

Table 40. Summary of results for vitamin B³ (niacinamide), mg/kg

a Identified as a non-representative or non-quantitative value; excluded from statistical analysis.

Fig. 29. Vitamin B³ (niacinamide) mass fraction as a function of production sequence
5.2.7. Vitamin B⁵ (Pantothenic Acid)

The manufacturer LC-MS/MS and HAMQAP [\[16\]](#page-41-0) measurement results for vitamin B_5 (pantothenic acid) are summarized in [Table 41.](#page-72-0) [Fig.](#page-72-1) 30 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer		HAMQAP Exercise 5								
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods		
1:1	77.0	E002	75.62	74.12	75.30	75.01	0.79	LC-MS/MS		
2:1	70.8	E005	61.00	60.00	60.00	60.33	0.58	LC-Abs or PDA		
3:1	74.7	E007	75.60	72.90	73.90	74.13	1.37	LC-Abs or PDA		
4:1	70.9	E010	71.07	75.98	78.10	75.05	3.60	LC-Abs or PDA		
5:1	74.1	E012	3.34 ^a	3.5 ^a	3.2 ^a			LC-Abs or PDA		
5:2	74.2	E013	$<$ 200 $^{\circ}$	$<$ 200 $^{\circ}$	$<$ 200 $^{\circ}$			LC-Abs or PDA		
6:1	77.5	E023	69.00	69.10	68.70	68.93	0.21	LC-MS/MS		
7:1	71.7	E030	78.10	73.20	75.30	75.53	2.46	LC-Abs or PDA		
8:1	69.1	E040	100 ^a	69.1^a	103.7°			LC-MS		
1:2	74.0	E042	72.73	69.58	70.62	70.98	1.60	LC-MS		
8:2	65.0	E047	70.00	70.00	70.00	70.00	0.00	LC-Abs or PDA		
N:	11				N:	8				
Mean:	72.6				Mean, Pooled SD: 71.25		1.75			
SD:	3.6				SD:	5.09				

Table 41. Summary of results for vitamin B⁵ (pantothenic acid), mg/kg

Fig. 30. Vitamin B⁵ (pantothenic acid) mass fraction as a function of production sequence

5.2.8. Vitamin B⁶ (Pyridoxine)

The manufacturer LC-FL and HAMQAP [\[16\]](#page-41-0) measurement results for vitamin B_6 (pyridoxine) are summarized in [Table 42.](#page-73-0) [Fig.](#page-73-1) 31 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 42. Summary of results for vitamin B⁶ (pyridoxine), mg/kg

Fig. 31. Vitamin B⁶ (pyridoxine) mass fraction as a function of production sequence

5.2.9. Biotin

The manufacturer LC-MS/MS and HAMQAP [\[17\]](#page-41-1) measurement results for biotin are summarized i[n Table 43.](#page-74-0) [Fig.](#page-74-1) 32 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 43. Summary of results for biotin, mg/kg

Fig. 32. Biotin mass fraction as a function of production sequence

5.2.10. Folic Acid

The manufacturer LC-MS/MS and FNSQAP [\[18\]](#page-41-2) measurement results for folic acid are summarized in [Table 44.](#page-75-0) [Fig.](#page-75-1) 33 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 44. Summary of results for folic acid, mg/kg

Fig. 33. Folic acid mass fraction as a function of production sequence

5.2.11. Vitamin B¹² (Cyanocobalamin)

The manufacturer LC-ICP-MS [\[14\]](#page-35-0) and NIST LC-ICP-MS measurement results for vitamin B_{12} (cyanocobalamin) are summarized in [Table 45.](#page-76-0) [Fig.](#page-76-1) 34 displays these results as an approximate function of the production sequence, as determined by the pallet and box number. The NIST measurement results for CNCbl in the SRM 1869 control material agreed well with the noncertified value stated in its COA and with those in previous LC-ICP-MS measurements.

Table 45. Summary of results for vitamin B¹² (cyanocobalamin), µg/kg

Fig. 34. Vitamin B¹² (cyanocobalamin) mass fraction as a function of production sequence

5.2.12. Vitamin C (Ascorbic Acid)

The manufacturer LC-Abs [30-32] and HAMQAP [\[17\]](#page-41-1) measurement results for vitamin C (ascorbic acid) are summarized in [Table 46.](#page-77-0) [Fig.](#page-77-1) 35 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 46. Summary of results for vitamin C (ascorbic acid), mg/kg

Fig. 35. Vitamin C (ascorbic acid) mass fraction as a function of production sequence

5.2.13. Vitamin D² (Ergocalciferol)

The manufacturer LC-MS/MS [33] and HAMQAP [\[16\]](#page-41-0) measurement results for vitamin D_2 (ergocalciferol) are summarized in [Table 47.](#page-78-0) [Fig.](#page-78-1) 36 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

	Manufacturer	HAMQAP Exercise 5								
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods		
1:1	0.1140	E002	0.95 ^a	1.1 ^a	1.29 ^a			LC-MS		
2:1	0.1143	E003	< 0.200 ^a	< 0.200 ^a	< 0.200 ^a			LC-MS/MS		
3:1	0.1168	E005	0.0525	0.0516	0.0533	0.0525	0.0009 LC-MS			
4:1	0.1203	E012	3 ^a	2.9 ^a	2.6 ^a			LC-Abs or PDA		
5:1	0.1178	E014	0.0660	0.0560	0.0580	0.0600		0.0053 LC-Abs or PDA		
5:2	0.1193	E015	2.28 ^a	1.39 ^a	2.19 ^a			LC-Abs or PDA		
6:1	0.1173	E023	0.1090	0.1070	0.1080	0.1080		0.0010 LC-MS/MS		
7:1	0.1145	E030	< 0.100 ^a	< 0.100 ^a	$<$ 0.100 $^{\circ}$			LC-MS/MS		
8:1	0.1133	E047	< 0.200 ^a	< 0.200 ^a	< 0.200 ^a			LC-Abs or PDA		
1:2	0.1123	E057	0.1140	0.1140	0.1220	0.1167		0.0046 LC-MS/MS		
8:2	0.1183				N:	4				
N:	11				Mean, Pooled SD:	0.0843	0.0036			
Mean:	0.1162				SD:	0.0327				
SD:	0.0026									

Table 47. Summary of results for vitamin D² (ergocalciferol), mg/kg

Fig. 36. Vitamin D² (ergocalciferol) mass fraction as a function of production sequence

5.2.14. Vitamin D³ (Cholecalciferol)

The manufacturer LC-MS/MS [\[33\]](#page-78-2) and HAMQAP [\[16\]](#page-41-0) measurement results for vitamin D_3 (cholecalciferol) are summarized in [Table 48.](#page-79-0) [Fig.](#page-79-1) 37 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

	Manufacturer			HAMQAP Exercise 5				
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods
1:1	0.1058	E002	0.0600	0.0700	0.0800	0.0700	0.0100	LC-MS
2:1	0.1063	E003	< 0.200 ^a	< 0.200 ^a	< 0.200 ^a			LC-MS/MS
3:1	0.1055	E005	0.0523	0.0511	0.0522	0.0519	0.0007	LC-MS
4:1	0.1060	E007	1.15^{a}	1.19 ^a	1.1^a			LC-Abs or PDA
5:1	0.1048	E012	2 ^a	2 ^a	2.2 ^a			LC-Abs or PDA
5:2	0.1050	E014	0.0470	0.0470	0.0520	0.0487	0.0029	lLC-Abs or PDA
6:1	0.1030	E015	1.5 ^a	2.12 ^a	1.48°			LC-Abs or PDA
7:1	0.1033	E023	0.1020	0.0998	0.1030	0.1016	0.0016	LC-MS/MS
8:1	0.1065	E030	$<$ 0.100 $^{\circ}$	$<$ 0.100 $^{\circ}$	< 0.100 ^a			LC-MS/MS
1:2	0.1073	E041	10.9 ^a	12.1^a	12.5^a			LC-Abs or PDA
8:2	0.1088	E047	0.1000	0.1000	0.1000	0.1000	0.0000	lLC-Abs or PDA
N:	11	E057	0.1020	0.1090	0.1000	0.1037	0.0047	LC-MS/MS
Mean:	0.1056				N:	6		
SD:	0.0017				Mean, Pooled SD:	0.0793	0.0047	
SD:								

Table 48. Summary of results for vitamin D³ (cholecalciferol), mg/kg

Fig. 37. Vitamin D³ (cholecalciferol) mass fraction as a function of production sequence

5.2.15. Vitamin E Acetate (α-Tocopheryl Acetate)

The manufacturer LC-Abs [\[29\]](#page-66-0) and HAMQAP [\[17\]](#page-41-1) measurement results for vitamin E acetate (αtocopheryl acetate) are summarized i[n Table 49.](#page-80-0) [Fig.](#page-80-1) 38 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer		HAMQAP Exercise 6									
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods			
1:1	164	F005	137.06	143.24	149.76	143.35	6.35	ILC-Abs or PDA			
2:1	158	F017	71.35	77.54	92.74	80.54	11.01	LC-Abs or PDA			
3:1	162	F034	179.00	192.00	195.00	188.67	8.50	ILC-Abs or PDA			
4:1	164	F039	232.00	243.00	249.00	241.33	8.62	ILC-FL			
5:1	164	F046	138.54	110.76	113.96	121.09	15.20	ILC-Abs or PDA			
5:2	162	F074	13.28 ^a	13.79 ^a	16.9 ^a			LC-Abs or PDA			
6:1	165	F075	151.00	156.00	153.00	153.33	2.52	LC-FL			
7:1	158	F079	132.00	111.00	50.00	97.67	42.59	LC-MS/MS			
8:1	160	F088	81.43	56.39	80.20	72.67	14.12	ILC-Abs			
1:2	161				N:	8					
8:2	160				Mean, Pooled SD:	137.3	17.9				
N:	11				SD:	57.3					
Mean:	161.6										
SD:	2.5										

Table 49. Summary of results for vitamin E acetate (α-tocopheryl acetate), mg/kg

Fig. 38. Vitamin E acetate (α-tocopheryl acetate) mass fraction as a function of production sequence

5.2.16. Free Vitamin E (Free α-Tocopherol)

The manufacturer LC-FL [\[29\]](#page-66-0) and HAMQAP [\[17\]](#page-41-1) measurement results for free vitamin E (free αtocopherol) are summarized in [Table 50.](#page-81-0) [Fig.](#page-81-1) 39 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer						HAMQAP Exercise 6			
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods	
1:1	61.0	F005	46.24	52.60	60.09	52.98	6.93	lLC-Abs or PDA	
2:1	59.4	F013	191 ^a	175 ^a	180°			LC-FL	
3:1	58.4	F017	26.36	26.81	29.86	27.68	1.90	lLC-Abs or PDA	
4:1	59.0	F030	188 ^a	185 ^a	185 ^a			LC-FL	
5:1	59.8	F033	203 ^a	200 ^a	199a			LC-Abs or PDA	
5:2	58.5	F034	71.40	66.30	71.20	69.63	2.89	lLC-Abs or PDA	
6:1	58.4	F039	211 ^a	221 ^a	227 ^a			LC-FL	
7:1	56.3	F046	23.36	22.94	22.66	22.99	0.35	LC-Abs or PDA	
8:1	56.5	F061	197.6°	202.1^a	213.1^a			LC-Abs	
1:2	60.5	F062	160.2 ^a	147.3°	140.5°			LC-FL	
8:2	54.5	F075	59.40	58.50	59.30	59.07	0.49	LC-FL	
N:	11	F088	52.80	43.77	53.93	50.17	5.57	LC-Abs	
Mean:	58.4				N:	6			
SD:	1.9				Mean, Pooled SD:	47.08	3.90		
					SD:	18.19			

Table 50. Summary of results for free vitamin E (free α-tocopherol), mg/kg

Fig. 39. Free vitamin E (free α-tocopherol) mass fraction as a function of production sequence

5.2.17. Total Vitamin E (Total α-Tocopherol)

The manufacturer LC-Abs/FL [\[29\]](#page-66-0) and HAMQAP [\[17\]](#page-41-1) measurement results for total vitamin E (total α-tocopherol) are summarized i[n Table 51.](#page-82-0) [Fig.](#page-82-1) 40 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer					HAMQAP Exercise 6					
ID	Value	Code	Rep ₁	Rep ₂	Rep ₃	Mean	SD	Methods		
1:1	210.4	F005	183.30	195.84	209.85	196.33	13.28	lLC-Abs or PDA		
2:1	203.4	F014	211.00	196.00	190.00	199.00	10.82	ILC-FL		
3:1	206.0	F017	26.36	26.81	29.86	27.68	1.90	ILC-Abs or PDA		
4:1	208.4	F020	193.00	206.00	204.00	201.00	7.00	ILC-FL		
5:1	209.2	F022	435 ^a	437 ^a	426 ^a			LC-Abs or PDA		
5:2	206.1	F031	212.92	215.66	225.79	218.12	6.78	lLC-Abs or PDA		
6:1	208.7	F033	364.00	361.00	356.00	360.33	4.04	ILC-Abs or PDA		
7:1	200.3	F046	149.59	123.86	126.50	133.32	14.15	ILC-Abs or PDA		
8:1	202.3	F069	0^a	0^a	0^a			LC-Abs or PDA		
1:2	207.2	F075	197.00	200.00	199.00	198.67	1.53	LC-FL		
8:2	200.3	F079	96.00	71.00	46.00	71.00	25.00	LC-MS/MS		
N:	11	F088	217.97	247.01	210.25	225.08	19.38	LC-Abs		
Mean:	205.7				N:	10				
SD:	3.6				Mean, Pooled SD:	183.1	12.7			
					SD:	91.0				

Table 51. Summary of results for total vitamin E (total α-tocopherol), mg/kg

Fig. 40. Total vitamin E (total α-tocopherol) mass fraction as a function of production sequence

5.2.18. β-Tocopherol

The manufacturer LC-FL [34-36] and HAMQAP [\[17\]](#page-41-1) measurement results for β-tocopherol are summarized in [Table 52.](#page-83-0) [Fig.](#page-83-1) 41 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 52. Summary of results for β-tocopherol, mg/kg

Fig. 41. β-Tocopherol mass fraction as a function of production sequence

5.2.19. γ-Tocopherol

The manufacturer LC-FL [\[34](#page-83-2)[-36\]](#page-83-3) and HAMQAP [\[17\]](#page-41-1) measurement results for γ-tocopherol are summarized in [Table 53.](#page-84-0) [Fig.](#page-84-1) 42 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 53. Summary of results for γ-tocopherol, mg/kg

Fig. 42. γ-Tocopherol mass fraction as a function of production sequence

5.2.20. δ-Tocopherol

The manufacturer LC-FL [\[34](#page-83-2)[-36\]](#page-83-3) and HAMQAP [\[17\]](#page-41-1) measurement results for δ-tocopherol are summarized in [Table 54.](#page-85-0) [Fig.](#page-85-1) 43 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 54. Summary of results for δ-tocopherol, mg/kg

Fig. 43. δ-Tocopherol mass fraction as a function of production sequence

5.2.21. Vitamin K (Phylloquinone)

The manufacturer LC-FL [37] and FNSQAP [\[18\]](#page-41-2) measurement results for vitamin K (phylloquinone) are summarized in [Table 55.](#page-86-0) [Fig.](#page-86-1) 44 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 44. Vitamin K (phylloquinone) mass fraction as a function of production sequence

5.2.22. β-Carotene

The manufacturer LC-Abs and FNSQAP [\[19\]](#page-41-3) measurement results for β-carotene are summarized i[n Table 56.](#page-87-0) [Fig.](#page-87-1) 45 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 56. Summary of results for β-carotene, mg/kg

Fig. 45. β-Carotene mass fraction as a function of production sequence

5.2.23. Lycopene

a

The manufacturer LC-Abs and FNSQAP [\[19\]](#page-41-3) measurement results for lycopene are summarized in [Table 57.](#page-88-0) [Fig.](#page-88-1) 46 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 57. Summary of results for lycopene, mg/kg

Fig. 46. Lycopene mass fraction as a function of production sequence

5.2.24. Lutein

The manufacturer LC-Abs and FNSQAP [\[19\]](#page-41-3) measurement results for lutein are summarized in [Table](#page-89-0) 58. [Fig.](#page-89-1) 47 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 58. Summary of results for lutein, mg/kg

Fig. 47. Lutein mass fraction as a function of production sequence

5.2.25. Carnitine

The manufacturer LC-MS/MS [38], NIST ID LC-MS [\[15\]](#page-38-0), and FNSQAP [\[19\]](#page-41-3) measurement results for carnitine are summarized in [Table 59.](#page-90-0) [Fig.](#page-90-1) 48 displays these results as an approximate function of the production sequence, as determined by the pallet and box number. The NIST measurement results for carnitine in the SRM 1849b control material agreed well with the noncertified value stated in its COA.

Table 59. Summary of results for carnitine, mg/kg

SD: 3.6

a

Fig. 48. Carnitine mass fraction as a function of production sequence

5.2.26. Choline

The manufacturer LC-MS/MS [\[38\]](#page-90-2), NIST ID LC-MS [\[15\]](#page-38-0), and FNSQAP [\[19\]](#page-41-3) measurement results for choline are summarized in [Table 60.](#page-91-0) [Fig.](#page-91-1) 49 displays these results as an approximate function of the production sequence, as determined by the pallet and box number. The NIST measurement results for choline in the SRM 1849b control material agreed well with the non-certified value stated in its COA.

Table 60. Summary of results for choline, mg/kg

Fig. 49. Choline mass fraction as a function of production sequence

5.2.27. *myo***-Inositol**

The manufacturer LC-PAD [39] results for *myo*-inositol are summarized in [Table 61.](#page-92-0) [Fig.](#page-92-1) 50 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 61. Summary of results *myo***-inositol, mg/kg**

Fig. 50. *myo***-Inositol mass fraction as a function of production sequence**

5.3. Proximates

5.3.1. Fat

The manufacturer base hydrolysis [40-43] and HAMQAP [\[17](#page-41-1)] measurement results for fat are summarized in [Table 62.](#page-93-0) [Fig.](#page-93-1) 51 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 51. Fat mass fraction as a function of production sequence

5.3.2. Protein

The manufacturer Kjeldahl [44] and HAMQAP [\[17\]](#page-41-1) measurement results for protein are summarized in [Table 63.](#page-94-0) [Fig.](#page-94-1) 52 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 63. Summary of results for protein, g/100 g

Fig. 52. Protein mass fraction as a function of production sequence

5.3.3. Carbohydrates

The manufacturer calculation and HAMQAP [\[17\]](#page-41-1) results for carbohydrates are summarized in [Table 64.](#page-95-0) [Fig.](#page-95-1) 53 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 64. Summary of results for carbohydrates, g/100 g

Fig. 53. Carbohydrates mass fraction as a function of production sequence

5.3.4. Solids

The manufacturer vacuum oven drying [45,46] and HAMQAP [\[17\]](#page-41-1) measurement results for solids are summarized in [Table 65.](#page-96-0) [Fig.](#page-96-1) 54 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 65. Summary of results for solids, g/100 g

Fig. 54. Solids mass fraction as a function of production sequence

5.3.5. Ash

The manufacturer dry ashing [47] and HAMQAP [\[17\]](#page-41-1) measurement results for ash are summarized in [Table 66.](#page-97-0) [Fig.](#page-97-1) 55 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 66. Summary of results for ash, g/100 g

Fig. 55. Ash mass fraction as a function of production sequence

5.3.6. Calories

The manufacturer calculation and HAMQAP [\[17\]](#page-41-1) results for calories are summarized in [Table 67.](#page-98-0) [Fig.](#page-98-1) 56 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 67. Summary of results for calories, kcal/100 g

Fig. 56. Calories as a function of production sequence

5.4. Cholesterol

The manufacturer GC-MS [48] measurement results for cholesterol are summarized in [Table 68.](#page-99-0) [Fig.](#page-99-1) 57 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 68. Cholesterol mass fraction results, mg/100 g

Fig. 57. Cholesterol mass fraction as a function of production sequence

Fatty Acids

5.5.1. Caproic Acid (C6:0)

The manufacturer GC-FID [49-53] measurement results for caproic acid (C6:0) are summarized in [Table 69.](#page-100-0) [Fig.](#page-100-1) 58 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 58. Caproic acid (C6:0) mass fraction as a function of production sequence

5.5.2. Caprylic Acid (C8:0)

The manufacturer GC-FID [\[49,](#page-100-2)[50\]](#page-100-3) measurement results for caprylic acid (C8:0) are summarized i[n Table 70.](#page-101-0) [Fig.](#page-101-1) 59 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 70. Caprylic acid (C8:0) mass fraction results, %

Fig. 59. Caprylic acid (C8:0) mass fraction as a function of production sequence

5.5.3. Capric Acid (C10:0)

The manufacturer GC-FID [\[49](#page-100-2)[,50\]](#page-100-3) measurement results for capric acid (C10:0) are summarized in [Table 71.](#page-102-0) [Fig.](#page-102-1) 60 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 71. Capric acid (C10:0) mass fraction results, %

Fig. 60. Capric acid (C10:0) mass fraction as a function of production sequence

5.5.4. Lauric Acid (C12:0)

The manufacturer GC-FID [\[49,](#page-100-2)[50\]](#page-100-3) measurement results for lauric acid (C12:0) are summarized in [Table 72.](#page-103-0) [Fig.](#page-103-1) 61 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 72. Lauric acid (C12:0) mass fraction results, %

Fig. 61. Lauric acid (C12:0) mass fraction as a function of production sequence

5.5.5. Myristic Acid (C14:0)

The manufacturer GC-FID [\[49,](#page-100-2)[50\]](#page-100-3) measurement results for myristic acid (C14:0) are summarized i[n Table 73.](#page-104-0) [Fig.](#page-104-1) 62 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 73. Myristic acid (C14:0) mass fraction results, %

Fig. 62. Myristic acid (C14:0) mass fraction as a function of production sequence

5.5.6. Palmitic Acid (C16:0)

The manufacturer GC-FID [\[49](#page-100-2)[,50\]](#page-100-3) measurement results for palmitic acid (C16:0) are summarized i[n Table 74.](#page-105-0) [Fig.](#page-105-1) 63 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 74. Palmitic acid (C16:0) mass fraction results, %

Fig. 63. Palmitic acid (C16:0) mass fraction as a function of production sequence

5.5.7. Palmitoleic Acid (C16:1)

The manufacturer GC-FID [\[49](#page-100-2)[,50\]](#page-100-3) measurement results for palmitoleic acid (C16:1) are summarized in [Table 75.](#page-106-0) [Fig.](#page-106-1) 64 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 75. Palmitoleic acid (C16:1) mass fraction results, %

Fig. 64. Palmitoleic acid (C16:1) mass fraction as a function of production sequence

5.5.8. Heptadecanoic Acid (C17:0)

The manufacturer GC-FID [\[49](#page-100-2)[,50\]](#page-100-3) measurement results for heptadecanoic acid (C17:0) are summarized in [Table 76.](#page-107-0) [Fig.](#page-107-1) 65 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 76. Heptadecanoic acid (C17:0) mass fraction results, %

Fig. 65. Heptadecanoic acid (C17:0) mass fraction as a function of production sequence
5.5.9. Stearic Acid (C18:0)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for stearic acid (C18:0) are summarized i[n Table 77.](#page-108-0) [Fig.](#page-108-1) 66 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 66. Stearic acid (C18:0) mass fraction as a function of production sequence

5.5.10. Oleic Acid (C18:1,9c)

The manufacturer GC-FID [\[49](#page-100-0)[,50\]](#page-100-1) measurement results for oleic acid (C18:1,9c) are summarized i[n Table 78.](#page-109-0) [Fig.](#page-109-1) 67 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 67. Oleic acid (C18:1,9c) mass fraction as a function of production sequence

5.5.11. Linoleic Acid (C18:2)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for linoleic acid (C18:2) are summarized i[n Table 79.](#page-110-0) [Fig.](#page-110-1) 68 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 79. Linoleic acid (C18:2) mass fraction results, %

Fig. 68. Linoleic acid (C18:2) mass fraction as a function of production sequence

5.5.12. -Linolenic Acid (C18:3,9c,12c,15c+9c,12c,15t)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for α -linolenic acid (C18:3,9c,12c,15c+9c,12c,15t) are summarized in [Table 80.](#page-111-0) [Fig.](#page-111-1) 69 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Manufacturer			
	ID	Value	
	1:1	0.5576	
	2:1	0.5538	
	3:1	0.5538	
	4:1	0.5566	
	5:1	0.5557	
	5:2	0.5547	
	6:1	0.5528	
	7:1	0.5538	
	8:1	0.5538	
	1:2	0.5566	
	8:2	0.5595	
N: 11			
	Mean:	0.5553	
	SD:	0.0021	

Table 80. -Linolenic acid (C18:3,9c,12c,15c+9c,12c,15t) mass fraction results, %

Fig. 69. -Linolenic acid (C18:3,9c,12c,15c+9c,12c,15t) mass fraction as a function of production sequence

5.5.13. -Linolenic Acid (C18:3)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for γ -linolenic acid (C18:3) are summarized in [Table 81.](#page-112-0) [Fig.](#page-112-1) 70 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 81. -Linolenic acid (C18:3) mass fraction results, %

Fig. 70. -Linolenic acid (C18:3) mass fraction as a function of production sequence

5.5.14. Arachidic Acid (C20:0)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for arachidic acid (C20:0) are summarized i[n Table 82.](#page-113-0) [Fig.](#page-113-1) 71 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 71. Arachidic acid (C20:0) mass fraction as a function of production sequence

5.5.15. Eicosenoic Acid (C20:1)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for eicosenoic acid (C20:1) are summarized in [Table 83.](#page-114-0) [Fig.](#page-114-1) 72 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 83. Eicosenoic acid (C20:1) mass fraction results, %

Fig. 72. Eicosenoic acid (C20:1) mass fraction as a function of production sequence

5.5.16. Homo--Linolenic Acid (C20:3,n6)

The manufacturer GC-FID [\[49](#page-100-0)[,50\]](#page-100-1) measurement results for homo- γ -linolenic acid (C20:3,n6) are summarized in [Table 84.](#page-115-0) [Fig.](#page-115-1) 73 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 84. Homo--linolenic acid (C20:3,n6) mass fraction results, %

Fig. 73. Homo--linolenic acid (C20:3,n6) mass fraction as a function of production sequence

5.5.17. Arachidonic Acid (C20:4,n6)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) and FNSQAP [\[19\]](#page-41-0) measurement results for arachidonic acid (C20:4,n6) are summarized in [Table 85.](#page-116-0) [Fig.](#page-116-1) 74 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 85. Arachidonic acid (C20:4,n6) mass fraction results, %

Fig. 74. Arachidonic acid (C20:4,n6) mass fraction as a function of production sequence

5.5.18. Behenic Acid (C22:0)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for behenic acid (C22:0) are summarized i[n Table 86.](#page-117-0) [Fig.](#page-117-1) 75 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 86. Behenic acid (C22:0) mass fraction results, %

Fig. 75. Behenic acid (C22:0) mass fraction as a function of production sequence

5.5.19. Docosahexaenoic Acid (C22:6, DHA)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) and FNSQAP [\[19\]](#page-41-0) measurement results for docosahexaenoic acid (C22:6, DHA) are summarized in [Table 87.](#page-118-0) [Fig.](#page-118-1) 76 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 87. Docosahexaenoic acid (C22:6, DHA) mass fraction results, %

Fig. 76. Docosahexaenoic acid (C22:6, DHA) mass fraction as a function of production sequence

5.5.20. Lignoceric Acid (C24:0)

The manufacturer GC-FID [\[49](#page-100-0)[,50\]](#page-100-1) measurement results for lignoceric acid (C24:0) are summarized in [Table 88.](#page-119-0) [Fig.](#page-119-1) 77 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 88. Lignoceric acid (C24:0) mass fraction results, %

Fig. 77. Lignoceric acid (C24:0) mass fraction as a function of production sequence

5.5.21. Nervonic Acid (C24:1)

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for nervonic acid (C24:1) are summarized i[n Table 89.](#page-120-0) [Fig.](#page-120-1) 78 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 89. Nervonic acid (C24:1) mass fraction results, %

Fig. 78. Nervonic acid (C24:1) mass fraction as a function of production sequence

5.5.22. Monounsaturated Fatty Acids

The manufacturer GC-FID [\[49](#page-100-0)[,50\]](#page-100-1) measurement results for monounsaturated fatty acids are summarized in [Table 90.](#page-121-0) [Fig.](#page-121-1) 79 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 90. Monounsaturated fatty acids mass fraction results, %

Fig. 79. Monounsaturated fatty acids mass fraction as a function of production sequence

5.5.23. Polyunsaturated Fatty Acids

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for polyunsaturated fatty acids are summarized in [Table 91.](#page-122-0) [Fig.](#page-122-1) 80 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 91. Polyunsaturated fatty acids mass fraction results, %

Fig. 80. Polyunsaturated fatty acids mass fraction as a function of production sequence

5.5.24. Saturated Fatty Acids

The manufacturer GC-FID [\[49](#page-100-0)[,50\]](#page-100-1) measurement results for saturated fatty acids are summarized i[n Table 92.](#page-123-0) [Fig.](#page-123-1) 81 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 92. Saturated fatty acids mass fraction results, %

Fig. 81. Saturated fatty acids mass fraction as a function of production sequence

5.5.25. Total *trans-***C18:1 and -C18:2 Fatty Acids**

The manufacturer GC-FID [\[49,](#page-100-0)[50\]](#page-100-1) measurement results for total *trans*-C18:1 and -C18:2 fatty acids are summarized in [Table 93.](#page-124-0) [Fig.](#page-124-1) 82 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Fig. 82. Total *trans***-C18:1 and -C18:2 fatty acids mass fraction as a function of production sequence**

5.5.26. Total Fatty Acids

The manufacturer GC-FID [\[49](#page-100-0)[,50\]](#page-100-1) measurement results for total fatty acids are summarized in [Table 94.](#page-125-0) [Fig.](#page-125-1) 83 displays these results as an approximate function of the production sequence, as determined by the pallet and box number. In addition to the fatty acids summarized in previous sections, the following fatty acids were assayed but not detected at a level of 0.01 g/100g: myristoleic acid (C14:1), pentadecanoic acid (C15:0), pentadecenoic acid (C15:1), heptadecenoic acid (C17:1), octadecatetraenoic acid (C18:4), eicosadienoic acid (C20:2), eicosatrienoic acid (C20:3,n3), arachidonic acid (C20:4,n3), eicosapentaenoic acid (C20:5), heneicosapentaenoic acid (C21:5), erucic acid (C22:1), docosadienoic acid (C22:2), docosatrienoic acid (C22:3), docosatetraenoic acid (C22:4), docosapentaenoic acid (C22:5,n3), and docosapentaenoic acid (C22:5,n6).

Fig. 83. Total fatty acids mass fraction as a function of production sequenceAmino Acids

5.6.1. Alanine

The manufacturer LC-Abs [54-57] measurement results for alanine are summarized in [Table 95.](#page-126-0) [Fig.](#page-126-1) 84 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 95. Alanine mass fraction results, g/100 g

Fig. 84. Alanine mass fraction as a function of production sequence

5.6.2. Arginine

The manufacturer LC-Abs [\[54-](#page-126-2)[57\]](#page-126-3) measurement results for arginine are summarized in [Table 96.](#page-127-0) [Fig.](#page-127-1) 85 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 96. Arginine mass fraction results, g/100 g

Fig. 85. Arginine mass fraction as a function of production sequence

5.6.3. Aspartic Acid

The manufacturer LC-Abs [\[54-](#page-126-2)[57\]](#page-126-3) measurement results for aspartic acid are summarized i[n Table](#page-128-0) [97.](#page-128-0) [Fig.](#page-128-1) 86 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 97. Aspartic acid mass fraction results, g/100 g

Fig. 86. Aspartic acid mass fraction as a function of production sequence

5.6.4. Cystine

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for cystine are summarized in [Table 98.](#page-129-0) [Fig.](#page-129-1) 87 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 98. Cystine mass fraction results, g/100 g

Fig. 87. Cystine mass fraction as a function of production sequence

5.6.5. Glutamic Acid

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for glutamic acid are summarized i[n Table](#page-130-0) [99.](#page-130-0) [Fig.](#page-130-1) 88 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 99. Glutamic acid mass fraction results, g/100 g

Fig. 88. Glutamic acid mass fraction as a function of production sequence

5.6.6. Glycine

The manufacturer LC-Abs [\[54-](#page-126-2)[57\]](#page-126-3) measurement results for glycine are summarized in [Table 100.](#page-131-0) [Fig.](#page-131-1) 89 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 100. Glycine mass fraction results, g/100 g

Fig. 89. Glycine mass fraction as a function of production sequence

5.6.7. Histidine

The manufacturer LC-Abs [\[54-](#page-126-2)[57\]](#page-126-3) measurement results for histidine are summarized i[n Table 101.](#page-132-0) [Fig.](#page-132-1) 90 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 101. Histidine mass fraction results, g/100 g

Fig. 90. Histidine mass fraction as a function of production sequence

5.6.8. Isoleucine

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for isoleucine are summarized in [Table](#page-133-0) [102.](#page-133-0) [Fig.](#page-133-1) 91 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 102. Isoleucine mass fraction results, g/100 g

Fig. 91. Isoleucine mass fraction as a function of production sequence

5.6.9. Leucine

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for leucine are summarized in [Table 103.](#page-134-0) [Fig.](#page-134-1) 92 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 103. Leucine mass fraction results, g/100 g

Fig. 92. Leucine mass fraction as a function of production sequence

5.6.10. Lysine

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for lysine are summarized in [Table 104.](#page-135-0) [Fig.](#page-135-1) 93 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 104. Lysine mass fraction results, g/100 g

Fig. 93. Lysine mass fraction as a function of production sequence

5.6.11. Methionine (Free)

The manufacturer LC-Abs [\[54,](#page-126-2)[55\]](#page-126-3) measurement results for methionine (free) are summarized in [Table 105.](#page-136-0) [Fig.](#page-136-1) 94 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 105. Methionine (free) mass fraction results, g/100 g

Fig. 94. Methionine (free) mass fraction as a function of production sequence

5.6.12. Methionine (Total)

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for methionine (total) are summarized in [Table 106.](#page-137-0) [Fig.](#page-137-1) 95 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 106. Methionine (total) mass fraction results, g/100 g

Fig. 95. Methionine (total) mass fraction as a function of production sequence

5.6.13. Phenylalanine

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for phenylalanine are summarized in [Table 107.](#page-138-0) [Fig.](#page-138-1) 96 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 107. Phenylalanine mass fraction results, g/100 g

Fig. 96. Phenylalanine mass fraction as a function of production sequence

5.6.14. Proline

The manufacturer LC-Abs [\[54-](#page-126-2)[57\]](#page-126-3) measurement results for proline are summarized i[n Table 108.](#page-139-0) [Fig.](#page-139-1) 97 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 108. Proline mass fraction results, g/100 g

Fig. 97. Proline mass fraction as a function of production sequence

5.6.15. Serine

The manufacturer LC-Abs [\[54-](#page-126-2)[57\]](#page-126-3) measurement results for serine are summarized in [Table 109.](#page-140-0) [Fig.](#page-140-1) 98 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 109. Serine mass fraction results, g/100 g

Fig. 98. Serine mass fraction as a function of production sequence

5.6.16. Taurine

Taurine, g/100 g

0.0350

The manufacturer LC-Abs [\[54,](#page-126-2)[55,57,](#page-126-3)58] measurement results for taurine are summarized i[n Table](#page-141-0) [110.](#page-141-0) [Fig.](#page-141-1) 99 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 110. Taurine mass fraction results, g/100 g

Fig. 99. Taurine mass fraction as a function of production sequence

 $\sqrt{4}$

Production Sequence, Pallet

3

 $\sqrt{2}$

 $\mathbf 1$

5

6

 $\overline{7}$

8

5.6.17. Threonine

The manufacturer LC-Abs [\[54](#page-126-2)[-57\]](#page-126-3) measurement results for threonine are summarized in [Table](#page-142-0) [111.](#page-142-0) [Fig.](#page-140-1) 98 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 111. Threonine mass fraction results, g/100 g

Fig. 100. Threonine mass fraction as a function of production sequence

5.6.18. Tryptophan

The manufacturer LC-Abs [\[54](#page-126-2)[,55,57,](#page-126-3)59] measurement results for tryptophan are summarized in [Table 112.](#page-143-0) [Fig.](#page-143-1) 101 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 112. Tryptophan mass fraction results, g/100 g

Fig. 101. Tryptophan mass fraction as a function of production sequence
5.6.19. Tyrosine

The manufacturer LC-Abs [\[54](#page-126-0)[-57\]](#page-126-1) measurement results for tyrosine are summarized in [Table 113.](#page-144-0) Fig. [102](#page-144-1) displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 113. Tyrosine mass fraction results, g/100 g

Fig. 102. Tyrosine mass fraction as a function of production sequence

5.6.20. Valine

The manufacturer LC-Abs [\[54-](#page-126-0)[57\]](#page-126-1) measurement results for valine are summarized in [Table 114.](#page-145-0) Fig. [103](#page-145-1) displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 114. Valine mass fraction results, g/100 g

Fig. 103. Valine mass fraction as a function of production sequence

5.7. Nucleotide Equivalents

5.7.1. Adenosine Monophosphate (*AMP***)**

The manufacturer LC-Abs measurement results for adenosine monophosphate (AMP) are summarized in [Table 115.](#page-146-0) [Fig.](#page-146-1) 104 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 115. Adenosine monophosphate (*AMP***) mass fraction results, mg/kg**

Fig. 104. Adenosine monophosphate (*AMP***) mass fraction as a function of production sequence**

5.7.2. Cytidine Monophosphate (*CMP***)**

The manufacturer LC-Abs measurement results for cytidine monophosphate (CMP) are summarized in [Table 116.](#page-147-0) [Fig.](#page-147-1) 105 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 116. Cytidine monophosphate (*CMP***) mass fraction results, mg/kg**

Fig. 105. Cytidine monophosphate (*CMP***) mass fraction as a function of production sequence**

5.7.3. Guanosine Monophosphate (*GMP***)**

The manufacturer LC-Abs measurement results for guanosine monophosphate (GMP) are summarized in [Table 117.](#page-148-0) [Fig.](#page-148-1) 106 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 117. Guanosine monophosphate (*GMP***) mass fraction results, mg/kg**

Fig. 106. Guanosine monophosphate (*GMP***) mass fraction as a function of production sequence**

5.7.4. Uridine Monophosphate (*UMP***)**

The manufacturer LC-Abs measurement results for uridine monophosphate (*UMP*) are summarized in [Table 118.](#page-149-0) [Fig.](#page-149-1) 107 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 118. Uridine monophosphate (*UMP***) mass fraction results, mg/kg**

Fig. 107. Uridine monophosphate (*UMP***) mass fraction as a function of production sequence**

5.7.5. Total Nucleotide Equivalents

The manufacturer LC-Abs measurement results for total nucleotide equivalents (without inosine monophosphate) are summarized in [Table 119.](#page-150-0) [Fig.](#page-150-1) 108 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 119. Total nucleotide equivalents^a mass fraction results, mg/kg

a Without inosine monophosphate (IMP)

Fig. 108. Total nucleotide equivalents mass fraction as a function of production sequence^a

Sugars

5.8.1. Galactooligosaccharides (GOS)

The manufacturer HPAEC-PAD measurement results for galactooligosaccharides (GOS) are summarized in [Table 120.](#page-151-0) [Fig.](#page-151-1) 109 displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 120. Galactooligosaccharides (GOS) mass fraction results, g/100 g

Fig. 109. Galactooligosaccharides (GOS) mass fraction as a function of production sequence

5.8.2. Lactose

The manufacturer GC-FID [60,61] measurement results for lactose are summarized in [Table 121.](#page-152-0) Fig. [110](#page-152-1) displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 121. Lactose mass fraction results, %

Fig. 110. Lactose mass fraction as a function of production sequence

5.8.3. Glucose

The manufacturer GC-FID [\[60](#page-152-2)[,61\]](#page-152-3) measurement results for glucose are summarized i[n Table 122.](#page-153-0) Fig. [111](#page-153-1) displays these results as an approximate function of the production sequence, as determined by the pallet and box number.

Table 122. Glucose Mass Fraction Results, %

Fig. 111. Glucose mass fraction as a function of production sequence

5.8.4. Total Sugars

The manufacturer GC-FID [\[60](#page-152-2)[,61\]](#page-152-3) measurement results for total sugars are summarized in [Table](#page-154-0) [123.](#page-154-0) [Fig.](#page-154-1) 112 displays these results as an approximate function of the production sequence, as determined by the pallet and box number. In addition to the sugars summarized in the previous sections, fructose, galactose, maltose, and sucrose were assayed but not detected at a level of 0.10 %.

Table 123. Total sugars mass fraction results, %

Fig. 112. Total sugars mass fraction as a function of production sequence

6. Value Assignment

Statistical Approaches

Statistical analysis of the data collected for the characterization of SRM 1849b was provided by the NIST Statistical Engineering Division.

6.1.1. Determination of Method Means and Uncertainties

Data for most analytes was provided by the material manufacturer. Data for some analytes was also collected using one or more NIST methods. For each analyte, the method mean for each method is the mean of the measurements available for that analyte using that method. The uncertainty of each such mean is the standard error of that mean.

For some fatty acids, all measurement data reported by the material manufacturer were identical. For these analytes, a Type B uncertainty related to instrument resolution [\[12\]](#page-32-0) was incorporated into the uncertainties for the method means.

6.1.2. Determination of Interlaboratory Study Means and Uncertainties

There are often very marked differences between the results from the different collaborative laboratories. The method estimate for each analyte is the weighted median of the individual laboratory means for that analyte, with weights determined using a Laplace random effects model [62]. For most analytes in this SRM, the weighted median is equal to or very close to the unweighted median of laboratory means. The uncertainty of the weighted median is estimated using a bootstrap procedure based on a Laplace random effects model for the betweenlaboratory and within-laboratory effects [\[12,](#page-32-0)[62](#page-155-0)–65].

6.1.3. Assignment of Values and Uncertainties

For each analyte, the assigned value is the unweighted mean of the method estimates available for that analyte. When the value is based on more than one method, the uncertainty of the combined mean is estimated using a bootstrap procedure based on a Gaussian random effects model for the between-method effects [\[12](#page-32-0)[,63](#page-155-1)–[65\]](#page-155-1). If only one method is available for an analyte, then that method estimate is the analyte estimate, with its corresponding expanded uncertainty.

Graphical analyses and analyses of variance with 5 % significance level were utilized for datasets that included box information to address issues of possible inhomogeneity of the SRM. No evidence of significant box effects was observed.

A number of outlier measurements were flagged by the analysts and omitted from the calculations. The deviance of these measurements from the others exceeded the usual variation, often differing by an order of magnitude or more. Other measurements may be questionable but could not be determined to be unrepresentative extreme outliers because of the sparseness and variation of the rest of the data.

For some analytes, interlaboratory comparison data was not included in the final value assignment (e.g., vitamin A palmitate, total vitamin A, vitamin D_2 , vitamin D_3). In these cases, analysts suspect bias in the data as a result of calibration, chromatographic peak identification, or final result calculation.

Certificate of Analysis

The results of the statistical analysis for all analytes are presented in the COA for SRM 1849b Infant/Adult Nutritional Formula I (milk-based). The most current version of the COA for SRM 1849b Infant/Adult Nutritional Formula I (milk-based) is available at [https://shop.nist.gov/ccrz__ProductDetails?sku=1849b.](https://shop.nist.gov/ccrz__ProductDetails?sku=1849b)

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NIST SP 260-233r1 August 2024

Appendix A. List of Abbreviations and Acronyms

AAS atomic absorption spectrometry

Abs absorbance spectroscopy

ANOVA analysis of variance

CNCbl cyanocobalamin (vitamin B12)

COA Certificate of Analysis

CVAAS cold vapor atomic absorption spectroscopy

df degrees of freedom

FL fluorescence spectroscopy

FNSQAP Food Nutrition and Safety Measurements Quality Assurance Program

GC-FID gas chromatography with flame ionization detection

HAMQAP Health Assessment Measurements Quality Assurance Program

HPAEC high-performance anion-exchange chromatography

ICP inductively coupled plasma

ILS interlaboratory study

IC-ECD ion chromatography with electrical conductivity detection

ISE ion selective electrode

LC liquid chromatography

MS mass spectrometry

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MS/MS tandem mass spectrometry

NAA neutron activation analysis

NIST National Institute of Standards and Technology

NMR nuclear magnetic resonance spectroscopy

OES optical emission spectrometry

PAD pulsed amperometric detection

PTFE polytetrafluoroethylene

q 1 -NMR quantitative proton nuclear magnetic resonance spectroscopy

RG/M

Roese-Gottlieb/Mojonnier. Acid Digestion with Ether Extraction (AOAC 986.25 & 945.48, 989.05)

RM

reference material

SI International System of Units

SPE

solid phase extraction

SRM

Standard Reference Material

TXRF total reflection X-ray fluorescence spectrometry

WDXRF

wavelength dispersive X-ray fluorescence spectrometry

Appendix B. Change Log

