

# Gedatolisib, a well-tolerated pan-PI3K/mTOR inhibitor, exhibits potent therapeutic effects on gynecological cancer models regardless of their PI3K pathway mutational status

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EXPANDING TREATMENT OPTIONS

Abstract 4928

## BACKGROUND

- The PI3K, AKT, and mTOR (PAM) pathway (Figure 1) is one of the most commonly activated oncogenic pathways in gynecological cancers, such as endometrial cancer (EC), ovarian cancer (OC), and cervical cancer (CC) (Table 1). Loss of PTEN function and PIK3CA activating mutations are especially frequent in EC (Table 1).
- Targeting the PAM pathway, in combination with other cooperative oncogenic pathways (e.g., the estrogen pathway), is a promising strategy for cancer treatment, including gynecological cancers<sup>2-4</sup>. Currently, the combination of everolimus, an mTORC1 inhibitor, and letrozole, an aromatase inhibitor, is an NCCN recommended regimen for patients with recurrent or metastatic endometrial EC<sup>5</sup>.
- Most PAM inhibitors (PAMI) selectively spare or weakly inhibit one or more key PAM pathway components, which can lead to drug resistance<sup>2</sup>. To minimize drug resistance, a more comprehensive inhibition of the PI3K isoforms and downstream mTOR complexes may be required.
- We hypothesized that gedatolisib, which potently inhibits all Class I PI3K isoforms, mTORC1, and mTORC2, can more effectively inhibit proliferation of EC and other gynecological cancer cells than other PAM inhibitors targeting individual PAM pathway components.

Figure 1. The PI3K, AKT, and mTOR (PAM) Pathway

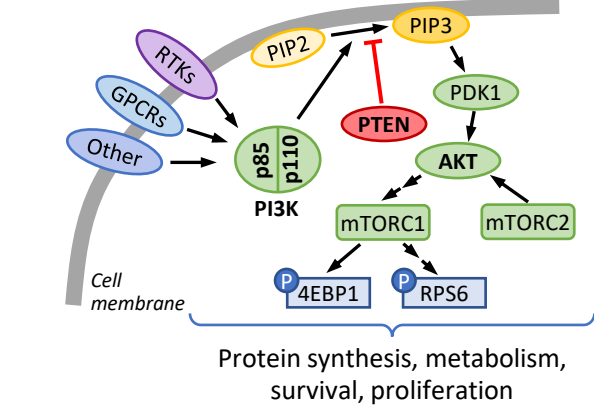


Table 1. Genetic Alterations of Key PAM Pathway Genes in EC, OC, and CC

	Endometrial Cancer (%)	Ovarian Cancer (%)	Cervical Cancer (%)
AKT1	5	4	4
AKT2	7	6	5
AKT3	9	7	2
PIK3CA	37	8	29
PIK3CB	11	8	8
PIK3R1	32	3	4
PIK3R2	7	6	1
PTEN	82	24	34

PIK3CA and PTEN values from Mills et al<sup>6</sup>; other values from cBioPortal analysis of the Cancer Genome Atlas (TCGA).

## METHODS

**Cell Lines.** A panel of 26 well-characterized EC, OC, and CC cell lines were used in this study (Table 2). Cells were maintained according to ATCC recommendations and authenticated by STR profiling. Genetic alterations in PAM pathway genes were identified by cBioPortal (https://www.cbioportal.org/) analysis of the Cancer Cell Line Encyclopedia (CCLE).

**Treatments with PAM Inhibitors.** Cells were seeded on 24- or 96-well plates coated with collagen-fibronectin-laminin, let attach overnight, and treated with PAM inhibitors (Table 3) at increasing concentrations to obtain dose response curves (DRCs). The seeding density of each cell line was optimized to ensure untreated cells remained in the growth phase throughout the assay.

**Viability and proliferation-normalized inhibition of growth rate (GR) assays.** After 72h PAMI treatment, cell viability was measured by RT-Glo MT assay (Promega) using a luminescence microplate reader. The normalized growth rate inhibition (GR) and per-division drug potency (GR50) metrics were calculated using the online GR calculator tool (http://www.grcalculator.org/grcalculator/) and the cell division time for each cell line. The normalized GR inhibition approach was used to rule out confounding effects of traditional IC50 metrics, such as the number of cell divisions occurring during the assay<sup>7-9</sup> (http://www.lincoproject.org).

**FACS analysis.** After 48h PAMI treatment, cells were harvested from 24-well plates, stained with a fixable viability dye (Zombie), fixed in 1.6% PFA, permeabilized with methanol, stained with antibodies, and analyzed by flow cytometry on the Agilent Novocyte 3005. To quantify cell proliferation, cells were cultured in the presence of 10 μM EdU for 2 hr prior to analysis by FACS using Click-iT chemistry (Invitrogen). Antibodies against pRPS6 and p4EBP1, two markers that integrate PAM signaling pathway outputs from PI3K/mTORC1 and mTORC2/pAKT (Figure 1), were used to assess PAM pathway inhibition by the PAMI<sup>10</sup>.

**Xenografts.** Immunodeficient female mice (nu/nu, SCID, or NOD.SCID) were inoculated subcutaneously in the flank with tumor pieces derived from MFE-280, MFE-296, AN3CA, or RL95-2 cell lines. When tumor size reached ~100-250 mm<sup>3</sup>, mice (N = 6-10 per arm) were randomly assigned to either a control vehicle group or a treatment group that received gedatolisib as indicated. Mice were treated every 4th day or twice a week for 12-21 days depending on the xenograft model.

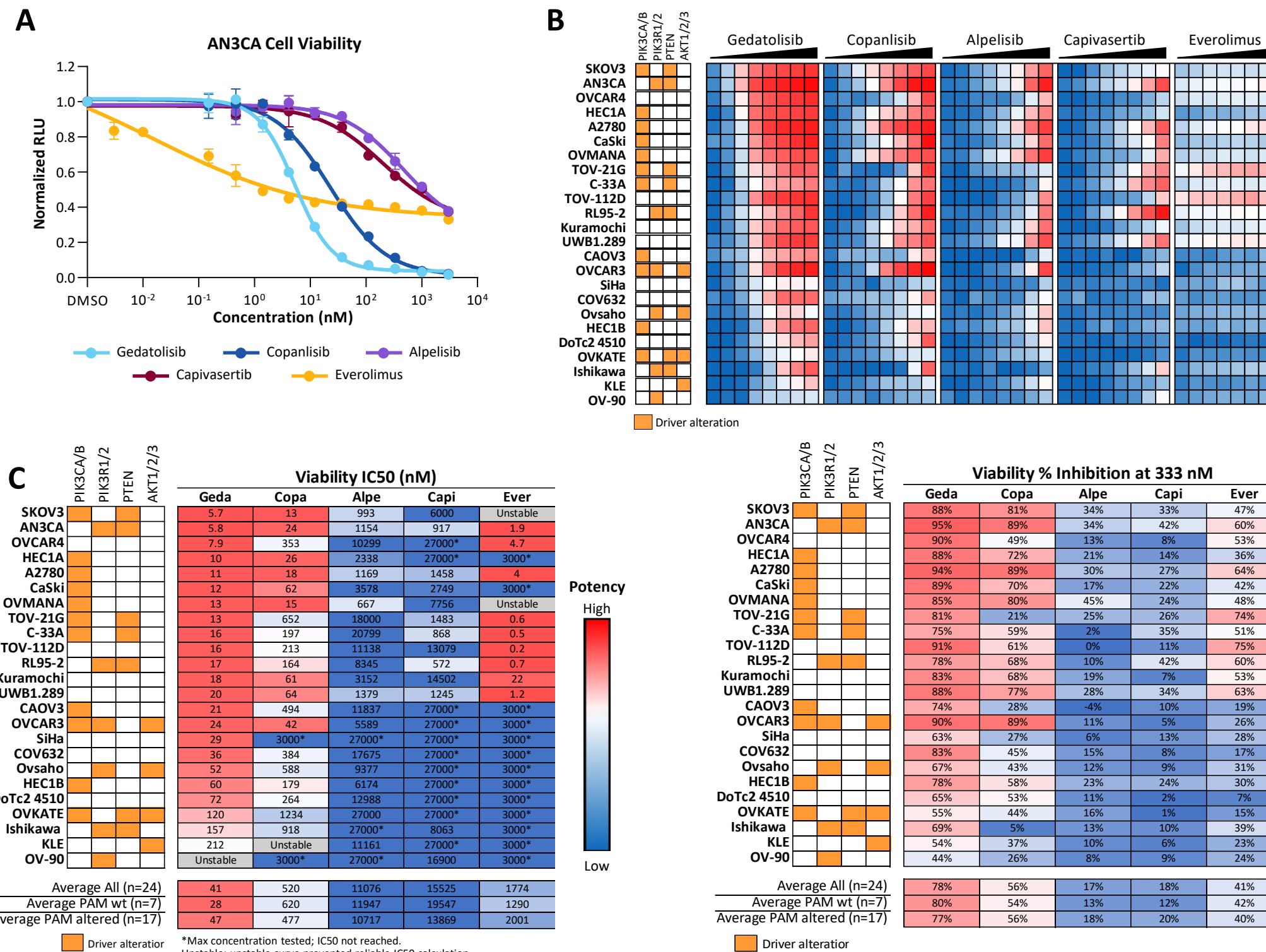
Table 2. Cancer Cell Lines Tested

Cell line	Cancer	PIK3CA	PIK3CB	PIK3R1	PIK3R2	PTEN	AKT1	AKT2	AKT3
AN3CA	EC	-	-	Mut	-	Mut	-	-	-
HEC1A	EC	Mut	-	-	-	-	-	-	-
HEC1B	EC	Mut, AMP	AMP	-	-	-	-	-	-
ISHIKAWA	EC	-	-	-	-	Mut	-	-	-
KLE	EC	-	-	-	-	-	-	AMP	AMP
MFE280	EC	Mut, AMP	-	-	-	-	AMP	AMP	-
MFE296	EC	Mut	-	-	-	Mut	-	-	-
RL95-2	EC	-	-	Mut	-	Mut	-	-	-
C33A	CC	Mut	-	-	-	Mut	-	-	-
CASKI	CC	Mut	-	-	-	-	-	-	-
DOT24510	CC	-	-	-	-	-	-	-	-
SIHA	CC	-	-	-	-	-	-	-	-
AZ780	OC	Mut	-	-	-	-	-	-	-
CAOV3	OC	Mut	AMP	-	-	-	-	-	-
COV632	OC	-	-	-	-	-	-	-	-
KURAMOCHI	OC	-	-	-	-	-	-	-	-
OV90	OC	-	-	-	-	HOMDEL	-	-	-
OVCA3	OC	-	AMP	Mut	-	-	-	AMP	-
OVCA4	OC	-	-	-	-	-	-	-	-
OVKATE	OC	Mut	-	-	-	HOMDEL	-	-	AMP
OVMANA	OC	Mut	-	-	-	-	-	-	-
OVSAHO	OC	-	-	-	-	AMP	-	-	AMP
SKOV3	OC	Mut, AMP	-	-	-	HOMDEL	-	-	-
TOV112D	OC	-	-	-	-	-	-	-	-
TOV21G	OC	Mut	-	-	-	Mut	-	-	-
UWB1289	OC	-	-	-	-	-	-	-	-

Table 3. PAM Inhibitors Tested

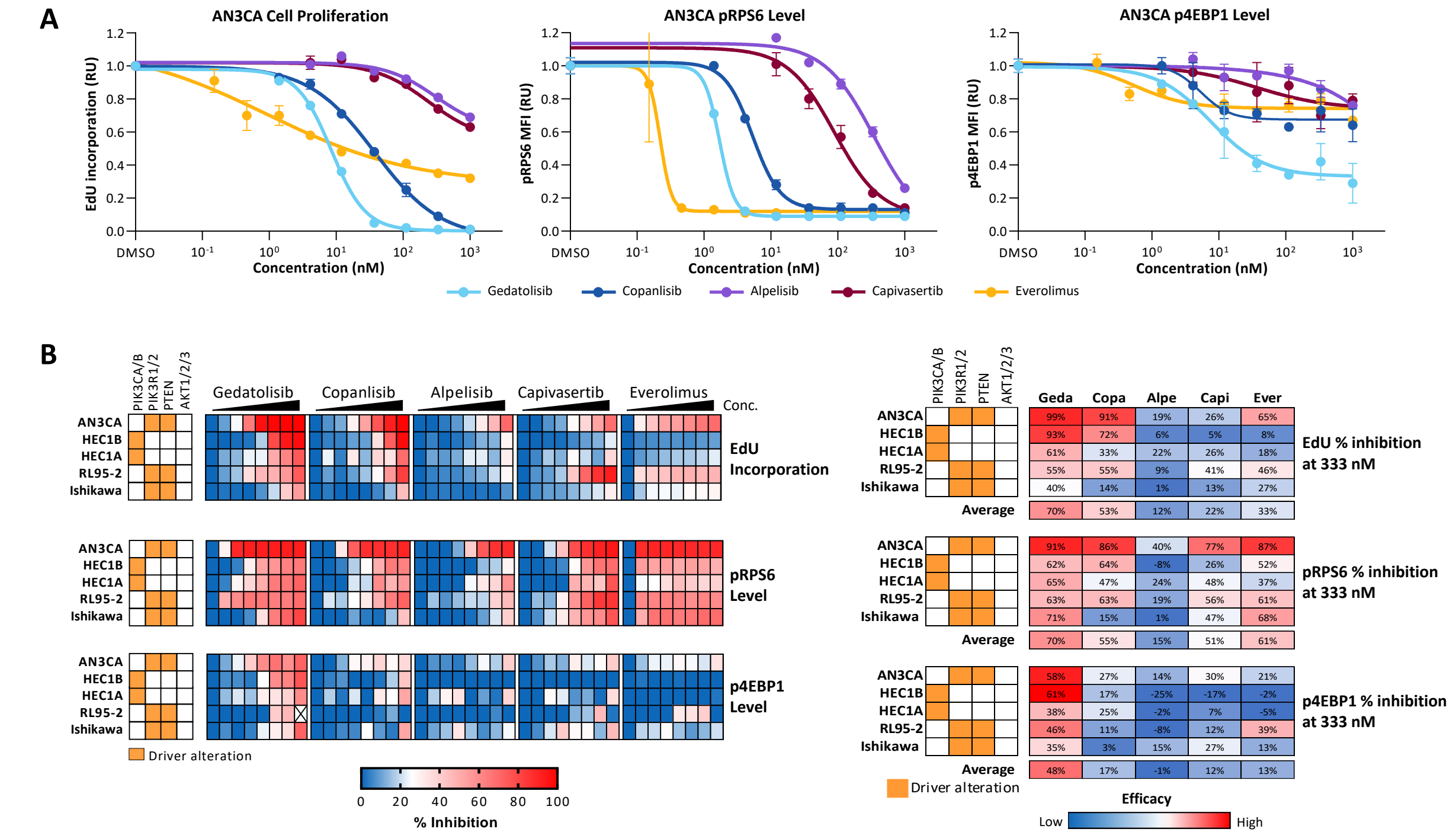
Drug	PAM specificity	Cell-free Assay Ki (nM)						
		PI3Kα	PI3Kβ	PI3Kγ	mTOR	AKT1	AKT2	AKT3
Gedatolisib	Pan-PI3K/mTOR	0.4	6	5.4	6	1.6	-	-
Copanlisib	Pan-PI3K	0.5	3.7	6.4	0.7	40	-	-
Alpelisib	PI3Kα	5	>1000	250	290	-	-	-
Capivasertib	AKT	-	-	-	-	-	3	8
Everolimus	mTOR	-	-	-	-	1.6	-	-

Figure 2. Cell Viability Analysis of EC, OC, and CC Cell Lines Treated With Gedatolisib and Other PAM Inhibitors



## RESULTS

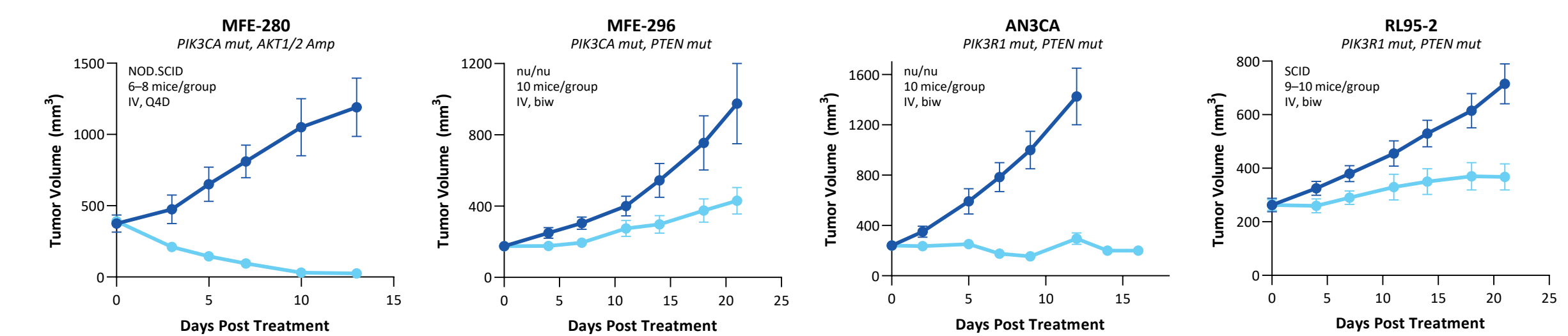
Figure 4. Assessment of DNA Replication and PAM Signaling Markers in EC Cell Lines



A panel of five EC cell lines was treated for 48 hours with escalating doses of PAM inhibitors and analyzed by flow cytometry for markers of DNA replication (EdU incorporation) and PAM signaling activity (pRPS6 and p4EBP1). (A) Examples of PAMI DRCs in AN3CA cells. Data are relative to DMSO-treated cells (set as 1) and represent mean +/- SD. (B) Heatmaps summarizing PAMI DRCs (1.4-1000 nM for gedatolisib, copanlisib, and everolimus; 12-9000 nM for alpelisib and capivasertib) in the five EC cell lines tested. (C) Table comparing the efficacy of gedatolisib and the other PAMI at inhibiting cell proliferation (EdU) and PAM signaling (pRPS6 and p4EBP1). The % inhibition shown in B and C is relative to DMSO-treated cells.

- The pan-PI3K/mTOR inhibitor, gedatolisib, suppressed DNA replication and PAM signaling in EC cells independent of their PAM pathway status more effectively than the other PAM inhibitors evaluated.

Figure 5. In Vivo Efficacy of Gedatolisib in Endometrial Cancer Xenograft Models



Immunodeficient mice were subcutaneously xenografted with MFE-280, MFE-296, AN3CA, or RL95-2 EC cell lines and treated with gedatolisib as described in the methods. Tumor growth inhibition (TGI) was assessed at the end of the treatment.

- In the MFE-280, MFE-296, AN3CA, or RL95-2 xenograft models, gedatolisib induced 145%, 68%, 95%, and 77% TGI respectively.

## SUMMARY AND CONCLUSIONS

- The pan-PI3K/mTOR inhibitor, gedatolisib, exhibited superior potency and efficacy across EC, OC, and CC cell lines with different PAM pathway mutational status *in vitro* relative to other PAM inhibitors.
- Potent anti-proliferative and cytotoxic effects of gedatolisib were seen in EC, OC, and CC cell lines regardless of PTEN, PI3K, or AKT mutational status.
- Gedatolisib demonstrated robust tumor growth inhibition *in vivo* in EC xenograft models with different PAM pathway mutational status.
- These findings provide a strong rationale to evaluate gedatolisib in gynecologic cancer clinical studies.

## References

- Mills SZ, et al. *JAMA Oncol.* 2016; 2(12): 1565-1573.
- Hafner M, et al. *Nat Rev Cancer.* 2015; 15: 7-24.
- Wang Q, et al. *Signal Transduct Target Ther.* 2020; 5(1): 137.
- Bregar A, et al. *Gynecol Oncol.* 2016; 140(2): 333-44.
- Slomovitz B, et al. *Gynecol Oncol.* 2022; 164(3):481-491.
- Ghandi M, et al. *Nature* 2019; 569(7757):503-508.
- Hafner M, et al. *Nat Methods.* 2016; 13 (6): 521-527.
- Hafner M, et al. *Curr Protoc Chem Biol.* 2017; 9 (2): 96-116.
- Niepel M, et al. *Curr Protoc Chem Biol.* 2017; 9 (2): 55-74.
- Josephs D, et al. *Transl Oncogenomics.* 2015; 7(Suppl 1): 33-49.